

## Therapeutic potential of convalescent plasma and hyperimmune immunoglobulins against SARS-CoV-2 BQ.1, BQ.1.1, and XBB variants

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Research Letter

COVID-19

Therapeutics

To the Editor: Convalescent plasma (CP) and hyperimmune intravenous immunoglobulins (IVIGs) are routinely used to treat patients with COVID-19. SARS-CoV-2 Omicron variants continue to evolve, generating multiple sublineages with increased transmissibility and antibody-escape mutations (1, 2). Several Omicron lineages that are currently circulating (BA.4, BA.5, BA.2.75, BA.2.75.2, BQ.1, BQ.1.1, and recombinant XBB) contain many mutations in spike protein (Supplemental Table 1; supplemental material available online with this article; <https://doi.org/10.1172/JCI168583DS1>), resulting in resistance to most therapeutic monoclonal antibodies as well as antibodies generated by SARS-CoV-2 vaccines (1, 2). Hyperimmune anti-SARS-CoV-2 IVIGs (hCoV-2IG) have been manufactured from pooled plasma units of hundred to thousands of convalescent individuals. hCoV-2IG contain immunoglobulin G at a 10-fold higher concentration compared with that in CP, and they are being evaluated for treatment of COVID-19 (3). To evaluate their therapeutic potential, 19 lots of hCoV-2IG prepared from convalescent individuals infected with SARS-CoV-2 in 2020, prior to circulation of Omicron; 20 IVIG preparations manufactured in 2019 (2019-IVIG) before the COVID-19 pandemic; and 8 IVIG lots manufactured from healthy plasma donations in 2020 (2020-IVIG) were analyzed for neutralization of SARS-CoV-2 Omicron BA.4/BA.5, BA.2.75, BA.2.75.2, BQ.1, BQ.1.1, and XBB subvariants in a pseudovirus neutralization assay (PsVNA) (Supplemental Methods). For comparison, we evaluated 8 CP samples from recovered patients with COVID-19 in early 2020 (2020-CP) and 9 CP samples from Omicron [...]

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# Therapeutic potential of convalescent plasma and hyperimmune immunoglobulins against SARS-CoV-2 BQ.1, BQ.1.1, and XBB variants

**To the Editor:** Convalescent plasma (CP) and hyperimmune intravenous immunoglobulins (IVIGs) are routinely used to treat patients with COVID-19. SARS-CoV-2 Omicron variants continue to evolve, generating multiple sublineages with increased transmissibility and antibody-escape mutations (1, 2). Several Omicron lineages that are currently circulating (BA.4, BA.5, BA.2.75, BA.2.75.2, BQ.1, BQ.1.1, and recombinant XBB) contain many mutations in spike protein (Supplemental Table 1; supplemental material available online with this article; <https://doi.org/10.1172/JCI168583DS1>), resulting in resistance to most therapeutic monoclonal antibodies as well as antibodies generated by SARS-CoV-2 vaccines (1, 2).

Hyperimmune anti-SARS-CoV-2 IVIGs (hCoV-2IG) have been manufactured from pooled plasma units of hundred to thousands of convalescent individuals. hCoV-2IG contain immunoglobulin G at a 10-fold higher concentration compared with that in CP, and they are being evaluated for treatment of COVID-19 (3).

To evaluate their therapeutic potential, 19 lots of hCoV-2IG prepared from convalescent individuals infected with SARS-CoV-2 in 2020, prior to circulation of Omicron; 20 IVIG preparations manufactured in 2019 (2019-IVIG) before the COVID-19 pandemic; and 8 IVIG lots manufactured from healthy plasma donations in 2020 (2020-IVIG) were analyzed for neutralization of SARS-CoV-2 Omicron BA.4/BA.5, BA.2.75, BA.2.75.2, BQ.1, BQ.1.1, and XBB subvariants in a pseudovirus neutralization assay (PsVNA) (Supplemental Methods). For comparison, we evaluated 8 CP samples from recovered patients with COVID-19 in early 2020 (2020-CP) and 9 CP samples from Omicron vaccine-breakthrough infections in 2022 (2022-CP).

2020-CP showed variable PsVNA50 titers against WA-1, ranging between 10 and 1,343 (geometric mean titer [GMT]: 154), but did not neutralize BQ.1, BQ.1.1, or XBB (Figure 1A and Supplemental Table 2). In contrast, 2022-CP demonstrated robust PsVNA50 titers against WA-1 (GMT: 926) and most neutralized BA.2.75, BA.2.75.2, and BA.4/BA.5 (GMT: 50, 59, and 71, respectively). However, only 4 2022-CP showed low neutralization of BQ.1 (GMT: 25), BQ.1.1 (GMT: 22), and XBB (GMT: 21).

As expected, the 2019-IVIG lots did not neutralize any SARS-CoV-2 strain. The 2020-IVIG lots (made from plasma units that were not screened for anti-SARS-CoV-2 neutralizing antibodies) had low PsVNA50 titers against WA-1 (GMT: 35) and did not neutralize Omicron variants (Figure 1A and Supplemental Table 2).

The 19 hCoV-2IG lots demonstrated robust neutralization of WA-1 (GMT: 1,615) (Figure 1A). Surprisingly, all 19 lots exhibited neutralization titers against BA.4/BA.5, ranging from 47 to 205 (GMT: 83). Importantly, 15 of the 19 hCoV-2IG lots also neutralized BA.2.75 and BA.2.75.2, with PsVNA50 titers of 22–430 (GMT: 37 and 32, respectively). At least 10 hCoV-2IG lots demonstrated presence of antibodies against BQ.1, BQ.1.1, and XBB subvariants, but the neutralization titers were further reduced (GMT: 21–25; Figure 1A and Supplemental Table 2). Strong correlations

were observed between PsVNA50 titers against WA-1/2020 and BA.4/BA.5, BA.2.75, and BA.2.75.2 ( $P < 0.0001$ ) for CP and hCoV-2IG (Figure 1B). In contrast, weak insignificant correlations were observed between PsVNA50 titers against WA-1/2020 and BQ.1, BQ.1.1, and XBB (Figure 1B).

Our study demonstrates that some hyperimmune COVID-IVIG lots manufactured in 2020 (2020-hCoV-2IG) neutralized several Omicron variants, similar to CP, from Omicron breakthrough infections in individuals with prior vaccination (2022-CP), at a level (PsVNA50 titer of  $>1:40$ ) predicted to provide protection against severe COVID-19 (4). Nevertheless, evolution of the variant landscape can increase resistance to antibodies elicited by prior SARS-CoV-2 infections and vaccination, especially against the newly emerged BQ.1, BQ.1.1, and XBB subvariants (5). Therefore, high-titer hCoV-2IG batches could be generated from donors who have been boosted recently with Omicron-containing bivalent vaccine and/or recovered from infection with Omicron following vaccination (hybrid immunity) (6). While there are logistical challenges to hyperimmune globulin production (e.g., long lead time), hCoV-2IG have notable advantages over CP, including standardization of dose, pathogen reduction, and measurements of anti-SARS-CoV-2 neutralizing titers prior to release. This could improve the hCoV-2IG therapeutic effectiveness against severe COVID-19 caused by circulating and emerging SARS-CoV-2 variants.

## Author contributions

SK and HG designed research. HG provided clinical specimens and unblinded clinical data. LB and SK performed assays. SK and HG contributed to manuscript writing.

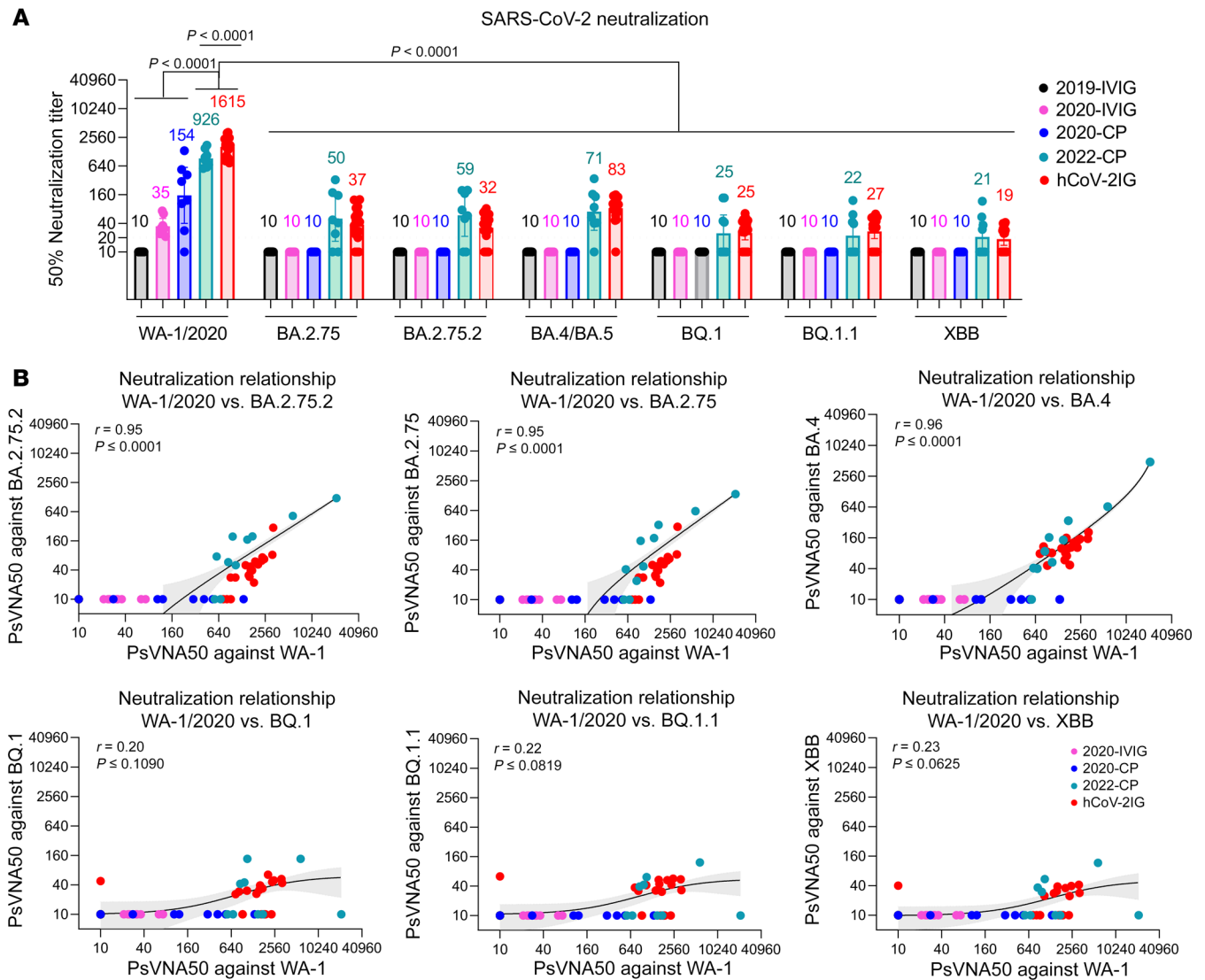
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**Figure 1. Neutralization of SARS-CoV-2 WA-1/2020 strain and Omicron subvariants by IVIG, convalescent plasma, and hCoV-2IG. (A)** SARS-CoV-2 neutralization assays were performed using pseudoviruses expressing the spike protein of WA-1/2020 or the Omicron subvariants in 293-ACE2-TMPRSS2 cells. SARS-CoV-2 neutralization titers were determined in each of the pre-pandemic 2019-IVIG ( $n = 20$ ; black), 2020-IVIG ( $n = 8$ ; pink), 2020 convalescent plasma (2020-CP;  $n = 8$ ; blue), 2022 convalescent plasma (2022-CP;  $n = 9$ ; turquoise), and hCoV-2IG ( $n = 19$ ; red) preparations. The assay was performed in duplicate to determine the 50% neutralization titer (PsVNA50). The heights of the bars and the numbers over the bars indicate the geometric mean titers, and the whiskers indicate 95% CIs. The horizontal dashed line indicates the limit of detection for the neutralization assay (PsVNA50 of 20). Differences between SARS-CoV-2 strains were analyzed by ordinary 1-way ANOVA, using Tukey's pairwise multiple-comparison test in GraphPad Prism version 9.3.1, and  $P$  values are shown. **(B)** Relationship of neutralizing antibodies against SARS-CoV-2 WA-1/2020 and Omicron subvariants. Correlation of SARS-CoV-2 WA-1/2020 neutralizing titer versus Omicron subvariant neutralizing titer for 2020-CP ( $n = 8$ ; blue), 2022-CP ( $n = 9$ ; turquoise), and hCoV-2IG ( $n = 19$ ; red). Correlations show Pearson's correlation coefficient ( $r$ ) and 2-tailed  $P$  values for all samples. The black lines in the scatter plots depict the linear fit of  $\log_2$ -transformed PsVNA50 values, with shaded area showing 95% CI.

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**Conflict of interest:** The authors have declared that no conflict of interest exists.

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