

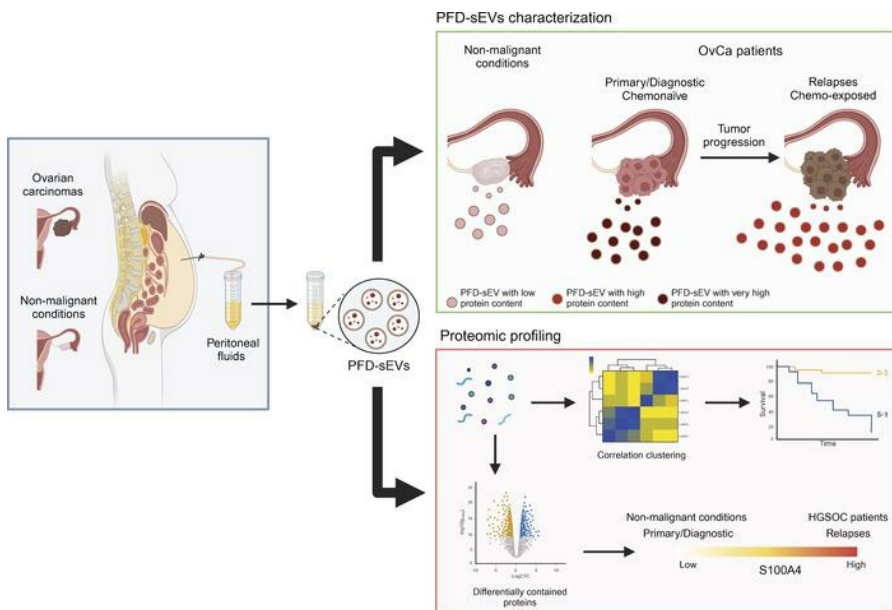
# Proteomic profiles of peritoneal-derived small extracellular vesicles correlate with outcome in ovarian cancer patients

Miguel Quiralte, ... , Sergio Ruiz-Llorente, Jesús García-Donas

*J Clin Invest.* 2024. <https://doi.org/10.1172/JCI176161>.

Research In-Press Preview Oncology

## Graphical abstract



Find the latest version:

<https://jci.me/176161/pdf>



## Title Page

**Manuscript title:** PROTEOMIC PROFILES OF PERITONEAL-DERIVED SMALL EXTRACELLULAR VESICLES CORRELATE WITH OUTCOME IN OVARIAN CANCER PATIENTS

## Authors and affiliations

Miguel Quiralte<sup>1,2</sup>, Arantzazu Barquín<sup>1,3</sup>, Mónica Yagüe<sup>1</sup>, Paloma Navarro<sup>1,2</sup>, Tatiana P. Grazioso<sup>1</sup>, Elena Sevillano<sup>1,3</sup>, Juan Francisco Rodriguez-Moreno<sup>1,3</sup>, Alejandra Balarezo-Saldivar<sup>1,2</sup>, Héctor Peinado<sup>4</sup>, Elena Izquierdo<sup>2</sup>, Carlos Millán<sup>5</sup>, Irene López<sup>5</sup>, Mario Prieto<sup>6</sup>, Rodrigo Madurga de Lacalle<sup>7</sup>, Ismael Fernández<sup>8</sup>, Sergio Ruiz-Llorente<sup>1,9,#</sup>, Jesús García-Donas<sup>1,2,3,#</sup>.

1. Laboratory of Innovation in Oncology HM CIOCC, MADRID (Clara Campal Comprehensive Cancer Centre), HM Sanchinarro University Hospital, 28050 Madrid, Spain.
2. Institute of Applied Molecular Medicine (IMMA), Faculty of Medicine, Universidad San Pablo-CEU, 28660 Boadilla del Monte, Madrid, Spain.
3. HM CIOCC (Clara Campal Comprehensive Cancer Centre), HM Sanchinarro University Hospital, 28050 Madrid, Spain.
4. Microenvironment and Metastasis Laboratory, Molecular Oncology Program, Spanish National Cancer Research Centre (CNIO), 28029 Madrid, Spain.
5. Gynecologic Unit, HM Montepríncipe University Hospital, 28660 Boadilla del Monte, Madrid, Spain.
6. Department of Pathological Anatomy - Therapeutic Targets Laboratory; HM Sanchinarro University Hospital, 28050 Madrid, Spain.
7. Faculty of Experimental Sciences, Universidad Francisco de Vitoria, 28223 Madrid, Spain.

8. R&D Oncology Business Unit, Pharmacogenomic & Cell Biology Departments, Pharmamar.
9. Department of Biomedicine and Biotechnology, Genetics Area, Universidad de Alcalá, 28805, Alcalá de Henares, Madrid, Spain.

**Co-Corresponding authors, #**

Jesus Garcia-Donas; <sup>1</sup>Laboratory of Innovation in Oncology HM CIOCC, MADRID (Clara Campal Comprehensive Cancer Centre), HM Sanchinarro University Hospital, 28050 Madrid, Spain. Phone: +34917567800; email address: jgarciadonas@hmhospitales.com

Sergio Ruiz Llorente; <sup>8</sup> Department of Biomedicine and Biotechnology, Genetics Area, Universidad de Alcalá. Phone: +34918854740; email address: sergio.ruizl@uah.es

**Conflict-of-interest statement:** The authors have declared that no conflict of interest exists.

## **TITLE**

Proteomic profiles of peritoneal-derived small extracellular vesicles correlate with outcome in ovarian cancer patients.

## **ABSTRACT**

Cancer-derived small extracellular vesicles (sEVs) are capable of modifying tumor microenvironment and promoting tumor progression. Ovarian cancer (OvCa) is a lethal malignancy that preferentially spreads through the abdominal cavity. Thus, the secretion of such vesicles into the peritoneal fluid could be a determinant factor in the dissemination and behavior of this disease. We designed a prospective observational study to assess the impact of peritoneal fluid-derived sEVs (PFD-sEVs) in OvCa clinical outcome. For this purpose, two patient cohorts were enrolled, including OvCa cases who underwent a diagnostic or cytoreductive surgery, and non-oncological patients as controls, who underwent abdominal surgery for benign gynecological conditions. PFD-sEVs systematic extraction from surgical samples enabled us to observe significant quantitative and qualitative differences associated with cancer diagnosis, disease stage and platinum chemosensitivity. Proteomic profiling of PFD-sEVs led to the identification of molecular pathways and proteins of interest and to the biological validation of S100A4 and STX5. In addition, unsupervised analysis of PFD-sEVs proteomic profiles in high-grade serous ovarian carcinomas (HGSOC) revealed two clusters with different outcomes in terms of overall survival. In conclusion, comprehensive characterization of the PFD-sEVs content provided a prognostic value with potential implications in HGSOC clinical management.

## INTRODUCTION

Ovarian cancer (OvCa) is the fifth leading cause of mortality among women and the foremost cause of death attributed to gynecological cancers in developed countries (1). OvCa patients present high recurrence rates within 16 to 22 months after completing radical surgery followed by conventional platinum-based chemotherapy (2). While numerous studies have assessed physical pelvic examination, transvaginal ultrasound, and serum carbohydrate antigen 125 (CA125) as screening methods, these procedures have shown low sensitivity and specificity (3–5). Thus, the majority of cases are diagnosed at advanced stages, when the tumor has typically disseminated throughout the abdominal cavity (6). Consequently, there is an urgent need to elucidate the molecular mechanisms triggering OvCa metastasis, identify predictive biomarkers, and discover novel targets to enhance therapeutic efficiency (7).

In the last decade, several studies have highlighted the role of extracellular vesicles (EVs) in the pathogenesis of multiple human diseases (8, 9). Small extracellular vesicles (sEVs) represent a subtype of EVs smaller than 200nm, that originate from multivesicular endosomes. These sEVs encapsulate a wide variety of biomolecules (including nucleic acids, proteins, and lipids, among others) that are secreted into the extracellular space and can modulate the physiology of target cells that internalize them (10). In the context of cancer, tumor-secreted sEVs contain molecules that are able to promote angiogenesis, facilitate intercellular communication between tumor cells and their microenvironment, modulate the immune response, and remodel surrounding and distal tissues. Consequently, sEVs could favor tumor progression through the establishment of a premetastatic niche (8, 11), the development of therapeutic resistance through a cross-talk between tumor cells, and immune evasion through the manipulation of immune cells (12, 13).

From a clinical perspective, the characterization of the cargo of sEVs secreted by the tumor could be relevant for the identification of diagnostic, predictive, and prognostic markers (8). Prior studies have isolated sEVs from peripheral blood and peritoneal fluids of patients with OvCa to identify diagnostic miRNA signatures (14–16). The protein content of OvCa-derived sEVs has been evaluated to a lesser extent and has been limited to the screening of specific proteins or the use of low-range customized panels for diagnostic purposes (16). However, these proteins may serve as the true effectors of most of the functions attributed to sEVs. Thus, high throughput proteomic analysis could help to determine and understand the role of sEVs and their cargo proteins in the outcome of OvCa patients.

The trans-coelomic route is the most frequent form of OvCa metastatic dissemination, being characterized by peritoneal spread and the formation of malignant ascites (17). Therefore, peritoneal fluid could act as a reservoir comprised of multiple cellular components and soluble factors composing the characteristic microenvironment of this tumor (15). Though massive ascites is a common event occurring at the latter steps of the disease, obtaining samples for proper analysis at the initial diagnosis stage is challenging (18). Considering these premises, we systematically collected peritoneal washings and malignant ascites during the surgical procedures undergone by 65 OvCa patients throughout the clinical course of their disease, as well as from 29 women who underwent abdominal interventions due to non-malignant conditions. The experimental approach subsequently included 1) the comparison of mass spectrometry proteomic profiling of the sEVs cargo in the OvCa cohort vs. the non-oncological patients cohort, 2) data integration with detailed clinical information available for OvCa patients, and 3) the biological validation of differentially sEVs-contained proteins through immunoblotting. Our work provides an essential proof of concept that the study of the protein cargo of PFD-sEVs represents an efficient strategy for the identification of prognostic signatures in ovarian cancer.

## RESULTS

### ***Clinical features and molecular characterization of OvCa and non-oncological patients.***

A prospective observational study was designed to recruit patients with OvCa and non-oncological gynecological conditions scheduled for abdominal surgery as part of the standard management. This study involved the participation of four health institutions in Madrid (Spain) which collaborated in the recruitment of participants.

From June 2018 to September 2022, a total of 74 peritoneal fluids derived from 65 OvCa patients (also mentioned in the text as OvCa cases) were collected. This cohort included serial samples from 8 patients, which comprised diagnostic laparoscopies and interval surgeries, with a third sample collected for one of these patients from a salvage surgery (in recurrence). Histological subtypes included high-grade serous adenocarcinoma (HGSOC, 54 cases [83.1%]), endometrioid carcinoma (6 [9.2%]), low-grade serous (2 [3.1%]), clear cell (2 [3.1%]) and mucinous (1 [1.5%]) (Table 1). Following the FIGO classification (19), tumor stage reported at the time of surgery included: stage I-II in 7 patients [10.8%], stage III in 38 [58.4%], and stage IV in 20 [30.8%] (Table 1). Next Generation Sequencing panels (FoundationOne CDx® and/or IdBRCA® test) were performed as routine practice in 54 patients, revealing *BRCA1/2* alterations in 16 patients (30%). Of these, 7 showed germline mutation (43.75%), 6 presented somatic mutation (37.5%), and the origin of the mutation could not be confirmed in 3 cases (Table 1). Detailed definitions pertaining to clinicopathological and molecular criteria can be found in the Methods section of the study.

In parallel, 29 non-oncological patients were recruited as the control cohort (hereinafter referred to as controls). The pathological diagnoses included: serous (n=7, [24.1%]) or mucinous (n=3, [10.35%]) ovarian cystadenomas; uterine myomas (n=7, [24.1%]),

ovarian cystic teratomas (n=2, [6.9%]), endometriosis (n=4, [13.8%]), normal tissue (prophylactic surgeries) (n=2, [6.9%]), uterine prolapse (n=3, [10.35%]), and endometrial hyperplasia (n=1, [3.5%]) (Table 1).

***Peritoneal fluid-derived sEVs (PFD-sEVs) concentration correlates with tumor stage and disease progression.***

Peritoneal fluid ultracentrifugation was used to extract sEVs from OvCa patients (n=74 samples from 65 patients) and control patients (n=29). Vesicle size analysis by Nanoparticle tracking analysis (NTA) in both cohorts showed an efficient and homogeneous collection of particles in the range of sEVs (<200nm; hereinafter referred to as PFD-sEVs) (Figure 1A) with similar average primary peak size for both sample sets (OvCa cases: 132.5nm vs. controls: 139.8nm) (Figure 1B). In accordance, transmission electron microscopy (TEM) analysis showed the presence of similar round-shaped vesicles in the sEVs size range (<200nm), both in samples from OvCa cases and controls (Figure 1C). NTA profiling additionally showed that there was no significant difference between the number of peritoneal-fluid derived particles obtained from OvCa cases when compared to controls (Figure 1D). However, a significant higher concentration was observed in OvCa cases when the size range was restricted to that associated with PFD-sEVs (OvCa cases,  $2.81 \times 10^{11}$  vs. controls,  $1.41 \times 10^{11}$  particles/ml (p/ml)) ( $p < 0.01$ ) (Figure 1E).

When OvCa cases were subdivided according to tumor stage, differences between OvCa cases and controls were shown to be mainly determined by a higher number of vesicles detected in patients with stages III-IV (Figure 1F). Notably, the HGSOc subtype was the only histology that showed statistically significant differences compared to controls. However, it is important to highlight that the number of OvCa cases with non-high-grade serous histology was limited, and the majority of them were diagnosed as stage I or II tumors (Supplemental Figure 1A). Thus, subsequent analyses concerning



the number of PFD-sEVs were restricted only to cases with HGSOC, the most frequently diagnosed histology both in clinical practice and within our study.

HGSOC-related analysis of particle concentration according to the surgical origin of the sample showed that the vesicular content was significantly higher upon relapse or after neoadjuvant chemotherapy (interval surgery) when compared to chemotherapy naïve samples (Figure 1G). Accordingly, when serial samples from patients were analyzed (n=8), the particle concentration tended to increase at interval surgeries (performed after neoadjuvant chemotherapy) when compared to diagnostic procedures (chemo-naïve samples) (2.0-fold change on average at interval) (Figure 1H). Regarding HGSOC platinum-based chemotherapy response, platinum-resistant cases showed a significantly higher PFD-sEVs concentration than platinum-sensitive cases (Figure 1I). However, no statistically significant differences in the concentration of sEVs were observed when HGSOC patients were classified according to the presence of BRCA1/2 alterations or HR status (Figure 1J). Finally, given that the presence of residual disease after cytoreductive surgery is one of the major predictive factors in ovarian cancer, we compared the quantity of sEVs in HGSOC patients who achieved an R0 resection vs. R1 and no significant differences were observed (Supplemental Figure 1B). Likewise, a lack of correlation was observed between the concentration of sEVs and the source of PFD-sEVs (peritoneal washings vs. ascites) (Supplemental Figure 1C).

***PFD-sEVs protein concentration is associated with HGSOC related clinical features.***

PFD-sEVs protein content was quantified through Bicinchoninic acid (BCA) assay. Protein content per particle (PCP) in PFD-sEVs was significantly higher (2.0-3.0 fold change) in OvCa cases diagnosed at stages III-IV (2.13 $\mu$ g per  $1\times 10^9$  particles) when compared to both controls (1.15 $\mu$ g per  $1\times 10^9$  particles) and low-stage tumors (0.75 $\mu$ g per  $1\times 10^9$  particles) ( $p$ -values: <0.01 and <0.05, respectively) (Figure 2A). It is worth

mentioning that the values corresponding to stages III-IV are 10 times higher than those observed by other authors in sEVs obtained from control patients serum (20), which may be related to an active secretory behavior of ovarian carcinoma and its tumor microenvironment at advanced stages. Similar to the quantitative data related to the number of vesicles (Figure 1B), we also detected significant differences in protein concentration when comparing HGSOC cases with controls, but no differences were observed for the remaining histological subtypes (Supplemental Figure 2A). However, as previously mentioned, non-HGSOC were poorly represented in our study and most cases were diagnosed at early stages, which could be a key confounding factor.

We subsequently analyzed PCP considering exclusively HGSOC histology. When these patients were classified based on surgical origin of the sample, it was noted that sEVs obtained from primary surgeries showed a higher PCP (3.5 $\mu$ g per  $1\times 10^9$  particles) than those acquired from interval (1.19 $\mu$ g per  $1\times 10^9$  particles) or relapse surgeries (1.71 $\mu$ g per  $1\times 10^9$  particles) ( $p$ -values:  $<0.001$  and  $<0.01$ , respectively) (Figure 2B). A similar trend was observed when comparing PCP from serial samples obtained from different HGSOC cases, particularly between diagnostic samples and interval surgery (6.5-fold change in concentration on average at diagnosis,  $p$ -values $<0.01$ ) (Figure 2C).

Given the patterns observed in HGSOC with respect to vesicle concentration (Figure 1G) and PCP (Figure 2B), we analyzed the potential association between these two variables through Pearson regression analysis and observed a significant inverse correlation (Supplemental Figure 2B). While interval and relapse surgeries showed a heterogeneous behavior, primary surgery samples homogeneously characterized by a reduced number of vesicles but increased protein content. Finally, while no statistically significant differences were noted regarding PCP when HGSOC patients were grouped either in R0 vs. R1 status (Supplemental Figure 2C), cases categorization based on the

sample source revealed a significant PCP increase in ascites vs. peritoneal washings (Supplemental Figure 2D).

**PDF-sEVs contain protein markers associated with the development of ovarian cancer.**

The purity of sEV preparations was confirmed through the analysis of exosomal markers (ALIX, TSG101, CD9) and negative protein markers (APOB and albumin), in compliance with the MISEV 2023 guidelines (21), in a representative subset of PFD-sEV samples obtained from OvCa cases or controls (Figure 2D). Moreover, immunoblotting profiling of the different supernatants or fractions collected throughout the ultracentrifugation-mediated extraction showed a robust expression of markers associated with exosome biogenesis (ALIX) or in its membrane dynamics and morphology (CD9 tetraspanin) exclusively in the PFD-sEVs suspension (Figure 2E). On the contrary, supernatants obtained in previous steps to the PFD-sEVs isolation (total peritoneal fluid or supernatant) displayed an absolute absence of expression for these markers (Figure 2E), denoting a selective and efficient extraction of PFD-sEVs.

To determine whether the PFD-sEVs fraction isolated from oncological patients contained vesicles specifically secreted by the ovarian tumor cells, we evaluated through immunoblotting the expression of the paired box 8 (PAX8) protein in a subset of OvCa cases (n=13) and controls (n=7). Despite the fact that PAX8 is mostly known for codifying a transcription factor essential in the physiology of thyroid follicular cells, its overexpression has been widely described in the context of ovarian carcinomas representing a reliable and widely used diagnostic marker for gynecological pathologies derived from the Fallopian tube secretory epithelial cells (22). As depicted in Figure 2F, all OvCa cases showed PAX8 expression in PFD-sEVs samples, whereas only 1 out of the 7 control samples tested showed positivity for this factor (normalized PAX8/CD9 ratio, p-value<0.001) (Figure 2F and Supplemental Figure 2E). These findings confirmed that

purified PFD-sEVs from OvCa cases were secreted by ovarian carcinoma cells prompting us to perform mass spectrometry profiling to identify protein biomarkers related to disease outcome, define molecular pathways modulated by tumor sEVs, and better classify HGSOC on the basis of their PFD-sEVs proteomic patterns.

***PFD-sEVs proteomics revealed a differential cargo of ovarian carcinoma-related biomarkers in patients cohort.***

The selection of OvCa cases and controls to be analyzed by mass spectrometry was performed considering the amount of total protein required for proteomic profiling ( $\geq 20\mu\text{g}$ ). The protein cargo of PFD-sEVs was profiled by liquid chromatography with tandem mass spectrometry (LC-MS/MS) in samples from 29 OvCa cases and 10 controls.

LC-MS/MS proteomic characterization of the PFD-sEVs cargo allowed the identification of 20899 peptides (FDR < 1%, calculated at peptide level) corresponding to 1825 proteins (Supplemental data 1). The list of proteins identified in the discovery cohort was compared against a reference list of exosome and extracellular vesicle markers obtained from reference repositories (Vesiclepedia and ExoCarta). As a result, we were able to detect the presence of more than 90% of the 100 most frequently proteins listed in both databases within our study samples, including the well-known sEVs markers CD9, CD63, and CD81 (Figure 3A). These findings, together with those related to the qualitative analysis of vesicle size by NTA, denoted that sEVs were efficiently isolated by the experimental approach used in our study.

PFD-sEVs proteomic data from HGSOC cases was further compared with previously published studies focused on identifying differentially expressed proteins between different specimens of OvCa cases vs. their corresponding normal counterparts (Supplemental Figure 3A) (23). Seven proteins (PEBP1, LGALS3, S100A8, FTL,

PSMA6, COL3A1 and AFM) showed significant changes ( $p$ -value $<0.05$ ) and a similar expression trend in our proteomic data when compared to such study (Figure 3B). Moreover, additional biomarkers presented a similar trend in terms of PFD-sEVs cargo when compared to the data previously described (Supplemental Figure 3B) (23). In accordance with the results obtained by Lai *et al.* by means of protein profiling in peripheral blood-derived exosomes, our data also confirmed the diagnostic value of FGG and APOA4, but not of MUC16 (which showed an opposite expression ratio) (Figure 3C). Paralleling the conclusion drawn from the PAX8 immunoblotting results, the robust correlation with these OvCa markers may suggest that our PFD-sEVs extracts represent biological specimens of interest for the study of the mechanisms involved in the progression and prognosis of HGSOE.

***Proteomic cargo differs depending on disease status and correlates with HGSOE overall survival.***

We then conducted an unsupervised clustering map based on the correlation of the 1825 proteins identified by mass spectrometry. The inclusion of samples from all OvCa cases and controls confirmed a separation of non-tumor specimens into two clusters with different expression profiles (cluster 1 and 2A), which included an OvCa sample (C1133). Interestingly, this patient presented an early-stage, low-grade endometrioid tumor, and underwent a microscopically-margin negative resection (R0) without the need for chemotherapy. With respect to the tumor samples, OvCa cases were subclassified into two well-differentiated clusters (cluster 2B-C vs. 3A-B) (Figure 4A; Supplemental data 1). In both clusters, we observed an indistinct distribution of HGSOE in two additional sets (subclusters 2B and 3A) and the preferential accumulation of non-HGSOE samples in specific branches (subclusters 2C and 3B). Cluster 2C included one patient who at the time of diagnosis had a mucinous cystadenoma (ctrl1351).

Next, we exclusively selected HGSOC cases, which represented the most frequent histology in our study, along with controls for further analysis (Figure 4B; Supplemental data 1). The cluster map exhibited a clear separation between cases and controls and two main clusters of OvCa cases (S-1 and S-2) were clearly distinguishable. The principal S-1 subcluster (subc. S-1) preferentially gathered samples obtained in surgeries from tumor recurrences (5/9 OvCa cases, 55%). These samples also clustered together in a specific subset with a robust correlation ratio (C1128, C467, C1445, and C424; correlation ratio average: 0.76). In contrast, the main S-2 subcluster (subc. S-2) preferentially included diagnostic and primary (chemo naïve) samples (8/10, 80%) and showed a subset of samples with the highest correlation rate among the entire analysis (C1083, C608, and C916; correlation ratio average: 0.82) harboring exclusively diagnostic specimens. Regarding the samples obtained from interval debulking surgeries, although their distribution was uneven in the clustering map, there was a tendency towards clustering close to each other (C1067, C618, and C469 or C660 and C416). It is worth mentioning that paired samples (C618, interval and C516, diagnostic) belonging to the same patient showed a high correlation in their vesicle content (correlation index: 0.72) and were grouped with samples obtained at similar surgical time points instead of being clustered together. Similar clustering results were obtained for both the comparison between recurrence and interval or primary surgery (Supplemental Figure 4, A and B; Supplemental data 1) and for the proteomic data-based Principal Component Analysis (PCA) (Supplemental Figure 4C; Supplemental data 1) which exclusively considered proteins detected for all the samples under study. These findings were corroborated by hierarchical clustering since ConsensusClusterPlus package analysis revealed a stable structure based on five clusters being able to differentiate two main clusters constituted by HGSOC samples (Supplemental Figure 4D; Supplemental data 1).

Considering the two main sets of correlations obtained from the proteomic data of the 23 HGSOC samples, significant differences were observed in terms of protein concentration (Figure 5A), which could denote a different secretory behavior depending on the disease status. Regarding patient's outcome, cluster S-1 (n=10) presented a tendency towards a shorter OS median (37.2 months) than S-2 (n=13) (not reached). Median overall follow-up was 34.4 months (27.7 months for the S-1 cluster vs 37.6 for the S-2 cluster) (Figure 5B). When comparison was restricted to HGSOC cases which samples were obtained by the time of the initial therapeutic intervention (diagnostic/primary and interval surgeries), this difference became significant (S-1 (n=5) vs S-2 (n=12); (median overall follow-up: 37.2 [29.7m for S-1 cluster vs 38.7months for S-2 cluster], [p- value<0.05]) (Figure 5B). Furthermore, despite the limited sample size and follow-up duration, significant differences in overall survival were observed when comparative analysis between S-1 vs S-2 was additionally restricted to patients undergoing primary or interval surgeries and harboring defects in HR, a molecular factor of relevance in the prognosis of HGSOC (Supplemental Figure 5). Globally, these results denote that the vesicle cargo does not only associates with HGSOC disease status and stage but also with its clinical outcome.

***Proteomics-based clusters of HGSOC cases are not associated with BRCA alterations, Homologous recombination status, or sensitivity to chemotherapy.***

It is widely established that around 20% of HGSOC cases harbor germline or somatic *BRCA* pathogenic alterations. However, this percentage rises up to 50% when mutations in other susceptibility genes associated to homologous recombination deficiency (HRD) are considered (19). Taking into account the genetic background of our HGSOC cases, pathogenic alterations impairing *BRCA1/2* or other HRD-related genes (such as *PALB2*, *BRIP1*, *RAD21*, *CCNE1* amplification) and HRD scores were similarly distributed among the proteomic clusters S-2 and S-1 (7/12 [58%] vs. 8/10 [80%], respectively) (Supplemental Figure 6A; Supplemental data 1). Moreover, platinum-based sensitivity

did not present any differences between the clusters (Supplemental Figure 6B; Supplemental data 1). Thus *BRCA*/HRD status and chemosensitivity, two well established prognostic factors, did not presented differences in distribution among PFD-sEVs proteomic clusters. This favors the hypothesis that the statistically significant difference regarding OS between S-1 and S-2 clusters could be attributed to their PFD-sEVs proteomic profiles rather than to any other confounding factor, especially when the comparison was restricted to non-relapsed samples. This restriction would avoid the selection bias of HGSOc cases that are considered suitable for a surgery at relapse (typically patients with low volume disease, good performance status, and long prior platinum free interval), and the biological alterations of the PFD-sEVs proteomic profiles induced by the exposition to chemotherapy (as previously shown). Unfortunately, the small numbers in our study preclude a multivariate analysis to definitively answer this intriguing question.

***Profiling of PFD-sEVs cargo proteins revealed factors associated with specific HGSOc clinical variables.***

The comparison of the proteomic profiling of HGSOc cases vs. controls revealed a differential PFD-sEVs cargo for 485 proteins (Figure 6A; Supplemental data 1) which represents more than 25% of the total amount of identified proteins. Among them, 96 different proteins appeared significantly over-contained (52/96) or under-contained (44/96) when stringent thresholds were considered ( $p$ -value<0.01 and z-scores>3).

Subsequent analysis performed for other clinically relevant variables, such as the degree of response to platinum-based combinations, *BRCA* status or the type of surgery which allowed PFD-sEVs collection, showed a differential content between the groups under study of 22, 21 and 181 proteins, respectively ( $p$ -value<0.05 and z-scores>2; Figure 6, B-D; Supplemental data 1). The higher number of factors deregulated for the surgical procedure comparison is in fact in accordance with the ability of the correlation clustering



analysis to distinguish among samples obtained at the diagnostic or primary intervention, after neoadjuvant chemotherapy or at recurrences. It is noteworthy that the String *in silico* analysis of proteins with differential content between HGSOC patients undergoing recurrent surgeries vs. primary or diagnostic procedures demonstrated an enrichment of factors related to the complement system and the S100A/ANXA protein families (Supplemental Figure 7A), which can also be observed in Figure 6D. As previously suggested, it also denotes that the ratio of cargo proteins responsible for the PFD-sEVs role in cancer progression is higher when compared to those related with the molecular basis of HGSOC pathogenesis or the chemoresistance.

***Validation studies supports a pro-tumorigenic role of S100A4 and STX5 derived from PFD-sEVs in HGSOC cases.***

As presented in Table 2, 25 of the identified proteins exhibited significant deregulation across various clinically relevant comparisons. Consequently, we established criteria for selecting targets of interest based on both their plausible biological role in tumor development and progression and expression ratios showing a consistent trend among the various comparisons under analysis. Considering these conditions, we directed our attention to two proteins among the 25 identified: S100A4 and STX5.

STX5 is actively involved in both autophagy events and in the transport between cellular compartments (ER to Golgi). Our dataset showed that *STX5* PFD-sEVs derived content was statistically higher both in HGSOC cases vs. controls (fold-change (FC): 4.55, adjusted *p*-value:0.008) and in HGSOC-platinum resistant vs. HGSOC-sensitive patients (FC: 2.27, adjusted *p*-value:0.015) (Supplemental data 1). Such significant differences were also observed when individual z-scores for *STX5* content were considered (Supplemental Figure 7B; Supplemental data 1). Regarding *S100A4* factor, extensive data supports its role in promoting the development and acquisition of aggressive phenotypes in different solid carcinomas (24, 25). In agreement, our proteomic profiling

showed that S100A4 PFD-sEVs content was significantly higher in HGSOC cases vs. controls (FC: 2.00, adjusted  $p$ -value<0.001) and in relapses vs. interval debulking or diagnostic specimens (FC: 2.57, adjusted  $p$ -value<0.0001) (Supplemental Figure 7, B and C; Supplemental data 1). Additional data supporting the potential role of S100A4 in HGSOC pathogenesis was the progressive increase of its z-scores throughout the clinical course of the disease (Supplemental Figure 7C) and its potential involvement with other functionally related proteins (ANXAs/S100A axis) (Supplemental Figure 7A).

Based on these assumptions, we performed a biological validation by Western-Blot on samples belonging to a new independent cohort of 22 HGSOC cases and 5 controls not included in the mass spectrometry analysis. Notably, the PFD-sEVs-content of both STX5 and S100A4 targets was significantly higher in HGSOC samples when compared to controls (STX5  $p$ -value=0.0098; S100A4  $p$ -value=0.008) (Figure 7, A and B, and Supplemental Figure 8A). In addition, subclassification of HGSOC cases according to the type of surgery confirmed a higher S100A4/CD9 ratio for any of the surgical-procedures with respect to controls, detecting a consistent trend with relapse samples ( $p$ =0.055) and showing robust differences when comparing to primary samples ( $p$ -value<0.01) (Supplemental Figure 8B). Due to the narrow nature of our dataset, there is a need to validate these observations in independent series; nevertheless, all these findings underline the potential role of PFD-sEVs contained proteins as factors involved in the development and progression of HGSOC.

***Functional enrichment analysis revealed molecular categories of interest: MTORC1 and Complement/Coagulation signaling pathways.***

*In silico* tools (*Enrichr* package or Gene Set Enrichment Analysis (GSEA)) were used to identify biological processes and functional categories altered among the differentially PFD-EVs contained proteins or between a rank ordered protein list, respectively. *ENRICH*R analysis of the data obtained from the comparison between HGSOC and

controls elucidated an enrichment of proteins associated with extracellular components such as vesicles, organelles and more specifically PFD-sEVs (Supplemental Figure 9A; Supplemental data 2), confirming the efficiency of our experimental approach and the robustness of our findings. Other relevant collections, such as the Molecular Signatures Database (MSigDB) Hallmarks and Reactome revealed an overrepresentation of proteins involved in complement and coagulation cascades, oxidative phosphorylation, immune events, and the phosphatidylinositol-3-kinase (PI3K) and the mammalian target of rapamycin (mTOR), and RAS signaling pathways (Supplemental Figure 9, B and C; Supplemental data 2).

GSEA analysis performed on the ranked lists based on proteomic quantifications for each comparison of interest (HGSOC cases vs. controls, resistant vs. sensitive, or recurrence vs. diagnostic) (Supplemental Figure 9D; Supplemental data 2) revealed a statistically significant enrichments in molecular categories similar to those defined by Enrichr. In this regard, we repeatedly observed associations with molecular classes related to the complement/coagulation pathway, cell to cell interaction (apical junctions) and motility (actin skeleton), estrogen mediated signaling, and oxidative phosphorylation. Collectively, these findings suggest that while certain individual proteins may actively promote HGSOC progression, it is possible that specific functional processes or pathways promoted by protein sets contained in PFD-sEVs may also play a pro-tumorigenic role.

***Pathways over and down represented in sEVs proteomic clusters from HGSOC cases could condition patients' clinical outcomes.***

PCA considering the 25 top proteins differentially up- or down- contained from the comparison of S-1 and S-2 clusters (Table 3) allowed to clearly distinguish HGSOC samples included in each subset (Supplemental Figure 10A), suggesting that such protein panel could represent a molecular signature set for the identification of patients

likely to have prolonged overall survival (OS). Worth mentioning that the correlation between our prognostic signature and other well-validated signatures based on HGSOC tissue (26) demonstrated that, despite being grounded in different omics tools and based on distinct biological materials, there was a certain degree of correlation between both studies. In this regard, out of the 126 genes belonging to the Yoshihara et al. signature, our study detected the expression of 8 (6.3%) of their corresponding proteins in all samples of our HGSOC cohort. Although only ANXA1 was included in our prognostic signature of 50 proteins, 6 of these 8 proteins (ANXA1, SERPINE1, APOL1, ALOX5AP, DSTN, and FCER1G, 75%) exhibited differential content in S1 vs S2 cluster comparison ( $p < 0.05$ ), with the seventh protein (PGK1) showing values close to statistical significance (Supplemental Figure 10B).

Notably, the Enrichr analysis revealed enrichment of functional categories related to epithelial differentiation in proteins overrepresented in the S-1 cluster, where samples from salvage surgeries at relapse tended to accumulate (Supplemental Figure 10C). On the other hand, and in line with what Yoshihara et al described (26), various categories related to immune processes were enriched among the overexpressed proteins in the S-2 cluster, where primary samples tended to cluster (Supplemental Figure 10D). These findings, which are in accordance with those observed in previous comparisons (HGSOC cases vs. controls and relapses vs. primary/diagnostic specimens, Reactome subset, Supplemental Figure 9, B and C), supports a key role of immune processes in HGSOC tumorigenesis and are in line with prior communications associating the activity of the immune system to a better clinical outcome (27).

## DISCUSSION

We present the results of a prospective observational study demonstrating how the characteristics of sEVs evolve dynamically during HGSOC progression and after exposure to platinum-based therapies. PFD-sEVs proteomic profiling classified HGSOC carcinomas into two groups with different clinical characteristics and overall survival, and independent of other well-established prognostic factors such as *BRCA* status or platinum sensitivity. These proteomic profiles identified immune processes as key features potentially modulated by PFD-sEVs. Taken together, our data suggest a role for PFD-sEVs in the initiation, progression, and clinical outcome of HGSOC, although their potential as disease biomarkers and therapeutic targets requires further investigation.

sEVs have been extensively studied in OvCa over the last decades (28). Many authors have conducted mechanistic analysis of their role in oncological processes such as cancer initiation, tumor dissemination, sensitivity to chemotherapy, and modulation of the tumor microenvironment (29, 30). However, these *in vitro* studies relied mostly on non-coding RNA and have had little translation into the clinic. Another relevant line of investigation focused on the assessment of the potential of serum sEVs as early diagnostic tools. In this regard, several microRNAs and microRNA signatures (14, 15, 31) and, to a lesser extent, sEVs proteins (16) have been detected in OvCa while being absent in healthy controls. However, as expected in a tumor with limited hematogenous spread, the proportion of tumoral sEVs is minimal and the correlation with clinical outcome was uncertain in most cases.

In our study, we collected peritoneal washings and malignant ascites from OvCa cases undergoing any oncologic surgery. This approach aimed to yield maximal amounts of tumor sEVs, enabling us to conduct high-throughput proteomic analysis of OvCa-derived vesicles. Additionally, we defined patterns of sEVs at different time points and evaluated

their evolution in serial samples. The preference for studying proteins over non-coding RNAs was based on their role as a true paracrine system linking tumor cells and their environment (32). Thus, our study evaluates the potential association between clinical features and outcome with tumor sEVs and their proteomic profiles in OvCa.

The characterization of the PFD-sEVs protein content within our OvCa cohort enabled us to confirm its malignant origin. Specifically, PAX8 expression, a marker of tumor cells (serous, endometrioid and clear cell OvCa) derived from the Fallopian tube secretory cell and less frequently expressed in benign pathologies (33, 34), was detected in all OvCa cases, but solely in one control.

Additionally, our study corroborates the results of previous proteomic studies in which differentially expressed proteins were determined in paired specimens from OvCa cases (tumor vs. normal tissue) or when comparing OvCa and control samples (urine and serum) (23). In this regard, we confirmed a significant overexpression of seven markers (PEBP1, LGALS3, S100A8, FTL, PSMA6, COL3A1 and AFM) when PFD-sEVs cargo from HGSOc cases vs. controls was compared. Minor differences observed for the remaining biomarkers could be attributable, not only to the different origin of the samples, but also to the contribution of the tumor microenvironment to the proteomic profile of OvCa-derived PFD-sEVs. In this sense, Lai *et al* recently proposed an OvCa-diagnostic signature based on the isolation of sEVs-derived serum markers (FGG, APOA4 and MUC16 (also known as CA125)) (16). Even though, we confirmed the diagnostic role of FGG and APOA4 in PFD-sEVs fraction, it was not the case for MUC16. Our results align with most prior clinical studies that have extensively tried to validate CA125 as a marker of ovarian cancer. Unfortunately, despite the large numbers of patients included in these studies, the specificity of this approach remains limited and must be interpreted alongside with additional diagnostic procedures (5, 35).

Our approach has also demonstrated that tumor stage, clinical progression, and the degree of exposition to chemotherapy strongly conditions both the number and protein content of PFD-sEVs in HGSOC, not only quantitatively but also qualitatively. In this sense, a significant association between tumor stage or progression and the amount of sEVs has been previously described in different types of solid tumors, such as melanoma and lung (15, 36). In the context of epithelial OvCa, it has been shown that different tumor-associated cell types (cancer cells, cancer-associated fibroblasts, immune component, among others) determine a high degree of heterogeneity in vesicle composition at the local and distal tumor environment (32), consequently promoting several pro-tumorigenic processes (37, 38). Such tumor-associated sEV heterogeneity would be consistent with the fact that our correlation clustering analyses based on PFD-sEVs profile classified HGSOC according to disease status, being able to discriminate paired samples depending on the studied timepoint. Concerning the benign pathologies within the control cohort, the various histological entities were distributed evenly across two clusters when compared to HGSOC cases. At this regard, it can be argued that the collected sEVs originated from non-pathological cells or abdominal structures, rather than from gynecological lesions. This observation is supported by the fact that PAX8 protein, a marker of gynecological pathologies, was isolated exclusively in one of the non-oncological controls.

Given that alterations in *BRCA* or HRD status do not substantially alter the differential profile of PFD-sEVs, it is worth noting that both tumor stage and progression and chemotherapy represent major forces of vesicle variability in HGSOC. Of particular interest are the results obtained from serial samples in which patients underwent two or more separate surgical interventions over time. Except for a single case that exhibited an opposite behavior, the number of vesicles increased, while the protein concentration per vesicle decreased after neoadjuvant treatment. Although this result is challenging to interpret, it is possible that the tumor cytolysis induced by chemotherapy led to this dual modification of sEVs-related features. Such hypothesis is supported by the fact that

physical (e.g., permeable vasculature, mechanical stress) and environmental conditions (acidic or hypoxic environments, cisplatin-based therapies, among others) in which tumor growth alter vesicle cargo and secretion ratios (32, 39, 40). However, considering that the patient displaying an opposite trend showed no differences in its clinical outcome, the significance of these changes should be explored in additional cohorts.

Accordingly, the study of these dynamic changes occurring in the content of PFD-sEVs may contribute to the identification of prognostic signature, as demonstrated by the significant correlation of their proteomic profile with OS in our cohort of HGSOc cases. Although based on different molecular approaches (transcriptomic vs. proteomic) and different biological samples (tumor tissue vs. PFD-sEVs), the correlation observed between the prognostic signature described by Yoshihara et al (26) and our proteomic data corroborates the prognostic potential of characterizing the content of PFD-sEVs in HGSOc. Of particular interest is the enrichment in categories related to the immune system, whose prognostic role was also addressed in the study by Yoshihara et al. Consistent with these findings, several OvCa studies have demonstrated how ascites-derived sEVs induce *in vitro* inactivation of CD3 and CD8 lymphocytes (41, 42) and activation of M2 macrophages (43) leading to disease progression. Globally our results not only confirm the role of these tumor-related sEVs in the modulation of the immune microenvironment, but also demonstrate their association with HGSOc-disease outcome. However, this potential should be further determined in future studies.

Detailed analysis of the hallmark-traits enriched along our proteomic data comparisons (HGSOc vs. controls, relapses vs. diagnostic/primary samples) revealed a recurrent association with categories such as the complement/coagulation cascade and immune responses, the PI3K/AKT/mTORC pathway, different metabolic processes, and hypoxia. The increased expression of complement factors (C4A, C4B, C5, C1R/S, among others) in PFD-sEVs between diagnostic samples and controls or recurrences samples could be related to previous findings suggesting that extracellular activation of this cascade by



TAMs represents an innate mechanism of immunosuppression that maintains a chronic inflammatory status promoting tumor progression (44, 45). Furthermore, it is worth noting the close relationship between peritoneal inflammation and the onset and progression of epithelial OvCa (46). On the other hand, oxidative phosphorylation (OXPHOS) and fatty acid processing were among the metabolic changes deregulated in our study. The involvement of these processes is consistent with the metabolic reprogramming that ovarian tumors may undergo, allowing them to prioritize oxidative phosphorylation over glycolysis to fuel tumor cells under hypoxic conditions (47). Notably, OvCa dependence on OXPHOS has been associated not only with increased survival (48) and proliferation of cancer-initiating stem cells, but also with increased chemosensitivity of tumor populations with high OXPHOS ratios (48), therefore, representing a promising therapeutic strategy. All these premises indicate that the PFD-sEVs content is a faithful reflection of the tumor progression and its intrinsic cell heterogeneity.

Considering previously published data regarding our set of deregulated proteins, we proceeded to the biological validation of proteins of interest (STX5 and S100A4) using immunoblotting as an alternative technique for protein-cargo evaluation. STX5 is a member of the SNARE protein family, which functions as an integral membrane protein playing a crucial role in autocrine and paracrine signaling through exocytosis and vesicle fusion-related events. Multiple oncogenic properties have been widely linked to the expression of SNARE proteins (49). Moreover, STX5 has been recently described to modulate PI3K/mTOR pathway activation, subsequently restraining cell adhesion and thus, favoring metastasis in hepatocellular carcinoma (50). Although STX5-mediated pro-tumorigenic mechanisms in HGSOc remain to be elucidated, our results demonstrate that its expression in PFD-sEVs is associated with tumor progression and, to a lesser extent, with the degree of response to chemotherapy which would be in line with analogous functions observed in other members of its protein family (49). Conversely, further literature links the overexpression of both S100A4 and other

members of its family to the development of metastasis through both intra- and extracellular functions. Several tumor components (cancer cells, cancer-associated fibroblasts or immune cells) are capable of secreting S100A4, highlighting that its protumorigenicity may be mediated by its inclusion in sEVs cargo (51, 52). Alluding to its involvement in OvCa development, several studies have described its role in the metastasis induction and chemoresistance (25, 53), and its potential use as a liquid prognostic marker for these neoplasms (54). In line with these assumptions, our results demonstrate that the oncogenic role of S100A4 in HGSOC is mediated, to some degree, by its secretion via PFD-sEVs and that it could serve as a biomarker of prognostic interest for HGSOC.

Our prospective study, which included massive proteomics on PFD-sEVs from OvCa patients, has allowed us to perform a longitudinal screening of the clinical course of the disease, demonstrating that the study of PFD-sEVs content can be useful in the identification of prognostic signatures. Restricting this analysis to the peritoneal cavity, we have further defined protumorigenic proteins and molecular pathways potentially involved in the intercellular communication of the tumor microenvironment, in line with the expected tumor heterogeneity of sEV content. Further studies in larger cohorts are required to validate the findings of this study.

## **METHODS**

### **Sex as a biological variable**

Our study exclusively focused on female samples as the disease under investigation (OvCa) is only relevant in females.

### **Clinical and molecular parameters for the categorization of OvCa patients**

Given that the OvCa patient samples were obtained from different types of surgeries, the nomenclature for these procedures is detailed in the following text: Primary: primary surgery performed as the initial therapeutic maneuver or diagnostic laparoscopy; Interval: Interval surgery post—neoadjuvant chemotherapy; Relapse: salvage surgery of recurrent disease. The surgical outcome terminologies are defined as follows: R0 is assigned when there is no residual macroscopic disease after surgery; R1 is attributed for any residual disease after surgery (regardless of the size). The term 'NA', "Not applicable" was used for those cases who underwent diagnostic laparoscopy procedures, where only a fragment of the tumor was excised for diagnosis purposes. Patient samples were classified based on the patient sensitivity to the next platinum-based chemotherapy administered after the collection of the sample; platinum sensitive patients were defined as those with a platinum-free interval (PFI), defined as the time from last chemotherapy to tumor relapse, >6 months, while platinum resistant patients were considered those with a PFI<6 months. *BRCA* status in patients was defined as follows: *BRCA* wild-type (WT), referring to the absence of pathogenic alterations in *BRCA1/2* and *BRCA* altered (ALT), indicating the existence of *BRCA* locus harboring a pathogenic alteration. Regarding the status of the homologous recombination (HR) pathway, HRD status reflects homologous recombination deficient-tumors, while HRP was used to denote those tumors with homologous recombination proficient features.

### **Sample collection, PFD-sEVs isolation and NTA profiling**

Immediately after the collection of peritoneal lavages from surgeries, centrifugation was performed at  $1500 \times g$  for 10 minutes to remove tissue debris and the supernatant was stored at  $-80^{\circ}\text{C}$ . Isolation and purification of sEVs from peritoneal fluid was systematically carried out using previously described protocols (21). Briefly, the samples were centrifuged at  $3,000 \times g$  for 20 min, followed by an additional centrifugation of the supernatant at  $12,000 \times g$  for 20 min in the Beckman Optima L-90K ultracentrifuge (Beckman Coulter, Fullerton, CA, USA) using the Type 55.2 rotor. sEVs were concentrated by centrifugation at  $100,000 \times g$  for 70 min. The resulting pellet was washed in 5ml PBS and collected through a second ultracentrifugation at  $100,000 \times g$  for 70 minutes using the SW55 rotor. The resulting pellet was resuspended in 100  $\mu\text{L}$  of PBS and the protein content was measured by bicinchoninic acid (BCA) assay (Pierce). Particle number was measured from an aliquot of 1-2  $\mu\text{l}$  of sEVs diluted in 1 ml of PBS using NTA (NanoSight; Malvern Panalytical Ltd. Worcestershire, United Kingdom) equipped with a violet laser (405 nm). For each sample, we recorded three 1-minute video clips.

### **Transmission Electron Microscopy (TEM)**

For negative staining, 5  $\mu\text{l}$  of the purified fractions resuspended in 2% PFA at a concentration of  $1 \times 10^{11}$  particles/ml were each deposited on a parafilm layer. A formvar/charcoal coated grid was placed over each drop and the sEVs were allowed to adsorb for 20 minutes. Then, they were washed over 5 drops of 100  $\mu\text{l}$  of PBS, 1 minute each and subsequently, each grid was placed over a drop of 50  $\mu\text{l}$  of 1% Glutaraldehyde in PBS and fixed for 5 minutes. After this, the grids were washed over 8 drops of 100  $\mu\text{l}$  of distilled water, 2 minutes each and placed on a drop of 50  $\mu\text{l}$  of Uranyl-Oxalate, contrasted for 5 minutes and left to dry. Finally, the grids were placed on a drop of 50  $\mu\text{l}$  of Methyl-Cellulose-Uranyl-Acetate for 10 minutes on ice and allowed to dry at room temperature. Visualization of the grids was performed on a JEOL JEM1010 transmission

electron microscope (100 KV). Images were recorded with a Gatan Orius 200 SC digital camera.

### **Immunoblotting**

sEV protein content was measured by BCA, and 10 µg aliquots were used for Western blot (WB) analysis. sEVs were lysed in Laemmli buffer at 96°C for 5 min to denature proteins and protein extracts were resolved by SDS-PAGE. The following antibodies were tested against Albumin (sc-271605, Santa Cruz Biotechnology, CA, USA), Apolipoprotein B (ab139401, Abcam, Cambridge, UK), TSG101 (ab125011, Abcam, Cambridge, UK), CD9 (ab58989, Abcam, Cambridge, UK) and ALIX (ab88743, Abcam, Cambridge, UK). Antibodies were chosen according to MISEV 2023 guidelines (21). Immunostaining against PAX8 (PA 0300; Biopat, Milan, Italy), a well-known ovarian carcinoma marker was also incorporated in the WB characterization panel. The biological validation employed the following antibodies against STX5 (HPA001358, Sigma-Aldrich, St. Louis, MO, USA) and S100A4 (ab93283, Abcam, Cambridge, UK). Clarity ECL Western Blotting Substrate (Biorad, Hercules, CA, USA) was used to visualize the bands and developed on the ChemiDoc imaging system (Biorad, Hercules, CA, USA). The intensity of the immunoreactive bands was quantified by densitometry using ImageJ (National Institutes of Health).

### **Proteomic analysis of PFD-sEVs by liquid chromatography–tandem mass spectrometry (LC-MS/MS) through differential analysis with TMT18-plex isobaric tagging**

For the proteomic analysis, LC-MS/MS tandem mass spectrometry was performed using an Ultimate 3000 Nano HPLC liquid nanochromatography system (Thermo Fisher Scientific, Waltham, MA, USA) coupled to an Orbitrap Exploris 240 mass spectrometer (Thermo Fisher Scientific, Waltham, MA, USA). Once the sEVs were isolated, they were concentrated on Nanosep columns (Omega, 10 kDa) with a starting volume greater than

50  $\mu$ l. They were then lysed with 4% SDS, followed by reduction and alkylation of cysteines using TCEP and MMTS, as reducing and alkylating reagent, respectively. A tryptic digestion is then performed on Protifi's commercial S-Trap columns. Peptide quantification (QuBit) is then carried out. Finally, isobaric tagging (TMT18-plex) is performed with 20  $\mu$ g of peptide mixture per condition. In this strategy, each sample is labelled with a different isotopic version of the reagent and combined into a single sample for mass spectrometric analysis.

The generated tryptic peptides are separated by liquid chromatography, desolvated and ionised before entering the Exploris 240 high-resolution mass spectrometer (with Orbitrap analyzer), where they are first detected as  $m/z$  ions in the range 375-1200  $m/z$ , and then the 20 most intense ones are taken from which MS/MS fragmentation spectra are obtained for identification. A long length gradient (120 min) is applied on a C18 column of 75 micrometer ID and 50 cm length. The different isotopic versions of the reagent are chemically identical, so that the same isotopically labelled peptide will generate the same precursor ion in the MS1 spectrum, differing only in the  $m/z$  values of the reporter or control ions characteristic for each of them. The relationship between the intensities of these control ions in the fragmentation spectra provides quantitative information at the peptide level, which in turn is related to differential expression levels at the protein level between samples.

### **LC-MS/MS Analysis and Data Extraction**

Proteins were identified in the raw files using the SEQUEST HT algorithm integrated into Proteome Discoverer 2.5 (Thermo Finnigan, Thermo Fisher Scientific, Waltham, MA, USA) (55). MS/MS scans were matched against UniProtKB human proteome database (2022\_06 Release) concatenated with the reverse decoy database obtained from the DecoyPYrat program (James Christopher Wright, Wellcome Trust Sanger Institute).

The parameters selected for the database searching as the following: trypsin digestion with two maximum missed cleavages allowed, precursor mass tolerance of 800 ppm (2 Da) and a fragment mass tolerance of 0.02ppm (0.02 Da) (56). The N-terminal and lysine TMT18-plex modifications, and the cysteine carbamidomethylation were chosen as fixed modifications, whereas the N-terminal acetylation, the methionine oxidation and Gln->pyro-Glu (N-term Q) were chosen as variable modification. The false discovery rate (FDR) was calculated using the refined method based on the results obtained by database searching. Quantitative information was extracted from the intensity of the TMT18-plex reporter ions in MS/MS spectra.

### **Bioinformatic analysis**

For comparative analysis of protein abundance changes, we used the Weighted Spectrum, Peptide, and Protein (WSPP) statistical model under the SanXoT software package (57). The model provides a detailed description of the behavior of technical variance, and by analyzing it independently at the spectrum, peptide, and protein levels, the model is able to capture separately the specific error sources of each SIL and MS method, demonstrating that error distributions are accurately modelled in all cases at the three levels. In addition, this model provides a standardized variable,  $Z_q$ , defined as the mean-corrected log<sub>2</sub>-ratio expressed in units of standard deviation at the protein level. ConsensusClusterPlus R package (58) was used to perform a hierarchical consensus clustering on protein expression data of control and HGSOC samples. A maximum of 6 clusters were fixed and the algorithm was trained over 10,000 repetitions using 80% of the samples. Pearson distance was used with complete hierarchical clustering. Results show a stable structure of 5 clusters (Supplemental Figure 4D), two of them belonging to HGSOC cases.

## **Systems Biology**

For the analysis of coordinated protein changes, we used both GSEA (59) and EnrichR (<https://maayanlab.cloud/Enrichr/>) *in silico* tools. GSEA calculates a normalized enrichment scores (NES) that measures the degree of enrichment and uses a permutation test to calculate a *p*-value that determines the statistical significance of the enrichment. The FDR (60) approach is used to control for multiple testing and reduce the likelihood of false positives. GSEA contains multiple molecular signature databases, including the ones used as Gene Ontology (GO), KEGG and Hallmarks. With respect to EnrichR, those differentially expressed proteins (adjusted *p*-value<0.05) for each comparison of interest were included in the analyses, which included relevant functional categories (Hallmarks, Reactome and Transcription Factor co-occurrence).

## **Statistical analysis**

GraphPad Prism (version 8.1.1, San Diego, CA, USA) was used to perform all statistical analyses. The following analyses, including the assessment of all particles and PFD-sEVs concentration, quantitative analysis of protein content per particle per 10<sup>9</sup> in PFD-sEVs, measurement of normalized ratios (PAX8/CD9, STX5/CD9, and S100A4/CD9) through immunoblotting in both control and OvCa samples, and comparisons of proteomic cargo between control and OvCa cases, were performed using the non-parametric Mann-Whitney test. For multiple comparisons *P*-values were adjusted using Benjamini-Hochberg method. For those analyses involving more than two groups, the Kruskal-Wallis test was employed, followed by Dunn's post hoc multiple comparison with Bonferroni's *P*-value adjustment. Results are expressed by Median and IQR. Analyses of PFD-sEVs concentration and protein content per particle per 10<sup>9</sup> in PFD-sEVs among the paired samples were conducted using the parametric paired t-test. Kaplan-Meier estimator and Cox method were performed for survival analyses. Different survival curves were compared using the log-rank test. In addition, the LIMMA package was used to determine differentially expressed proteins between OvCa specific clinical variables



and controls. For statistical significance, a  $p$ -value of less than 0.05 was considered significant.

### **Study approval**

The study protocol (21.01.1289E1-GHM) for sample collection was approved by the institutional ethics committee and patients provided written consent before being included in the study.

### **Data availability**

Every value of the data points depicted in the graphs of the manuscript is accurately reflected in the Supporting Data Values XLS file. Additionally, proteomic and differentially contained proteins data are available in the Supplemental data 1 XLS file. For the Enrichr and GSEA analyses (Supplemental Figures 9 and 10, C and D), data values are provided in the Supplemental data 2 XLS file.

## **Authors contributions**

MQ, SRL and JGD designed the research studies. All experiments and acquiring data were done by MQ, AB, MY, PN, ABS, SRL. Histological assessments were performed by MP. Human clinical sample collection and patient assessments were performed by AB, ES, JFR, CM, IL and JGD. Proteomic profiling data were analyzed by MQ, SRL, RM and IF. Reviewed data, provided experimental advice, and contributed to manuscript editing and revision were carried out by HP and EI. The manuscript was written by MQ, TG, SRL and JGD. The project was conceived and supervised by SRL and JGD. All authors reviewed the manuscript and provided feedback on it.

## BIBLIOGRAPHY

1. Redondo A, et al. SEOM clinical guideline in ovarian cancer (2020). *Clin Transl Oncol*. 2021;23(5):961–968.
2. Yamamoto CM, et al. Comparison of benign peritoneal fluid- and ovarian cancer ascites-derived extracellular vesicle RNA biomarkers. *J Ovarian Res*. 2018;11(1):20.
3. Jacobs IJ, et al. Ovarian cancer screening and mortality in the UK Collaborative Trial of Ovarian Cancer Screening (UKCTOCS): a randomised controlled trial. *The Lancet*. 2016;387(10022):945–956.
4. Chang L, et al. Liquid biopsy in ovarian cancer: recent advances in circulating extracellular vesicle detection for early diagnosis and monitoring progression. *Theranostics*. 2019;9(14):4130–4140.
5. Buys SS. Effect of Screening on Ovarian Cancer Mortality: The Prostate, Lung, Colorectal and Ovarian (PLCO) Cancer Screening Randomized Controlled Trial. *JAMA*. 2011;305(22):2295.
6. Torre LA, et al. Ovarian cancer statistics, 2018: Ovarian Cancer Statistics, 2018. *CA: A Cancer Journal for Clinicians*. 2018;68(4):284–296.
7. Feng W, et al. Exosomes promote pre-metastatic niche formation in ovarian cancer. *Mol Cancer*. 2019;18(1):124.
8. Becker A, et al. Extracellular Vesicles in Cancer: Cell-to-Cell Mediators of Metastasis. *Cancer Cell*. 2016;30(6):836–848.
9. Olejarz W, et al. Tumor-Derived Exosomes in Immunosuppression and Immunotherapy. *Journal of Immunology Research*. 2020;2020:1–11.

10. Raposo G, Stoorvogel W. Extracellular vesicles: Exosomes, microvesicles, and friends. *Journal of Cell Biology*. 2013;200(4):373–383.
11. Hoshino A, et al. Tumour exosome integrins determine organotropic metastasis. *Nature*. 2015;527(7578):329–335.
12. Ciardiello C, et al. Focus on Extracellular Vesicles: New Frontiers of Cell-to-Cell Communication in Cancer. *IJMS*. 2016;17(2):175.
13. van der Pol E, et al. Classification, Functions, and Clinical Relevance of Extracellular Vesicles. *Pharmacol Rev*. 2012;64(3):676–705.
14. Meng X, et al. Diagnostic and prognostic relevance of circulating exosomal miR-373, miR-200a, miR-200b and miR-200c in patients with epithelial ovarian cancer. *Oncotarget*. 2016;7(13):16923–16935.
15. Wang W, et al. Integrated analysis of ascites and plasma extracellular vesicles identifies a miRNA-based diagnostic signature in ovarian cancer. *Cancer Letters*. 2022;542:215735.
16. Lai H, et al. Protein Panel of Serum-Derived Small Extracellular Vesicles for the Screening and Diagnosis of Epithelial Ovarian Cancer. *Cancers*. 2022;14(15):3719.
17. Tan DS, Agarwal R, Kaye SB. Mechanisms of transcoelomic metastasis in ovarian cancer. *The Lancet Oncology*. 2006;7(11):925–934.
18. Rickard BP, et al. Malignant Ascites in Ovarian Cancer: Cellular, Acellular, and Biophysical Determinants of Molecular Characteristics and Therapy Response. *Cancers (Basel)*. 2021;13(17):4318.
19. González-Martín A, et al. Newly diagnosed and relapsed epithelial ovarian cancer: ESMO Clinical Practice Guideline for diagnosis, treatment and follow-up. *Annals of Oncology*. 2023;S0923753423007974.

20. Brennan K, et al. A comparison of methods for the isolation and separation of extracellular vesicles from protein and lipid particles in human serum. *Sci Rep.* 2020;10(1):1039.
21. Welsh JA, et al. Minimal information for studies of extracellular vesicles (MISEV2023): From basic to advanced approaches. *J of Extracellular Vesicle.* 2024;13(2):e12404.
22. Di Palma T, Zannini M. PAX8 as a Potential Target for Ovarian Cancer: What We Know so Far. *OTT.* 2022;Volume 15:1273–1280.
23. Toss A, et al. Ovarian Cancer: Can Proteomics Give New Insights for Therapy and Diagnosis? *IJMS.* 2013;14(4):8271–8290.
24. Kim B, et al. The role of S100A4 for bone metastasis in prostate cancer cells. *BMC Cancer.* 2021;21(1):137.
25. Hiruta A, et al. S100A4/Nonmuscle Myosin IIA/p53 Axis Contributes to Aggressive Features in Ovarian High-Grade Serous Carcinoma. *The American Journal of Pathology.* 2020;190(11):2304–2316.
26. Yoshihara K, et al. High-risk ovarian cancer based on 126-gene expression signature is uniquely characterized by downregulation of antigen presentation pathway. *Clin Cancer Res.* 2012;18(5):1374–1385.
27. Zhang L, et al. Intratumoral T cells, recurrence, and survival in epithelial ovarian cancer. *N Engl J Med.* 2003;348(3):203–213.
28. Wang K-H, Ding D-C. The Role and Applications of Exosomes in Gynecological Cancer: A Review. *Cell Transplant.* 2023;32:9636897231195240.

29. Li G, et al. Tumour-derived exosomal piR-25783 promotes omental metastasis of ovarian carcinoma by inducing the fibroblast to myofibroblast transition. *Oncogene*. 2023;42(6):421–433.
30. Wang X, Jiang L, Liu Q. miR-18a-5p derived from mesenchymal stem cells-extracellular vesicles inhibits ovarian cancer cell proliferation, migration, invasion, and chemotherapy resistance. *J Transl Med*. 2022;20(1):258.
31. Chen L, et al. Plasma exosomal miR-1260a, miR-7977 and miR-192-5p as diagnostic biomarkers in epithelial ovarian cancer. *Future Oncol*. 2022;18(26):2919–2931.
32. Lee AH, Koh IL, Dawson MR. The role of exosome heterogeneity in epithelial ovarian cancer. *Advances in Cancer Biology - Metastasis*. 2022;4:100040.
33. Kim J, et al. Diagnostic roles of PAX8 immunohistochemistry in ovarian tumors. *Pathol Res Pract*. 2023;250:154822.
34. Wang Y, et al. PAX8: a sensitive and specific marker to identify cancer cells of ovarian origin for patients prior to neoadjuvant chemotherapy. *J Hematol Oncol*. 2013;6:60.
35. Menon U, et al. Ovarian cancer population screening and mortality after long-term follow-up in the UK Collaborative Trial of Ovarian Cancer Screening (UKCTOCS): a randomised controlled trial. *Lancet*. 2021;397(10290):2182–2193.
36. Peinado H, et al. Melanoma exosomes educate bone marrow progenitor cells toward a pro-metastatic phenotype through MET. *Nat Med*. 2012;18(6):883–891.
37. Li W, et al. TGF $\beta$ 1 in fibroblasts-derived exosomes promotes epithelial-mesenchymal transition of ovarian cancer cells. *Oncotarget*. 2017;8(56):96035–96047.
38. Nieman KM, et al. Adipocytes promote ovarian cancer metastasis and provide energy for rapid tumor growth. *Nat Med*. 2011;17(11):1498–1503.

39. Samuel P, et al. Cisplatin induces the release of extracellular vesicles from ovarian cancer cells that can induce invasiveness and drug resistance in bystander cells. *Phil Trans R Soc B*. 2018;373(1737):20170065.
40. Bandari SK, et al. Chemotherapy induces secretion of exosomes loaded with heparanase that degrades extracellular matrix and impacts tumor and host cell behavior. *Matrix Biology*. 2018;65:104–118.
41. Asare-Werehene M, et al. Plasma Gelsolin Inhibits CD8+ T-cell Function and Regulates Glutathione Production to Confer Chemoresistance in Ovarian Cancer. *Cancer Res*. 2020;80(18):3959–3971.
42. Shenoy GN, et al. Sialic Acid-Dependent Inhibition of T Cells by Exosomal Ganglioside GD3 in Ovarian Tumor Microenvironments. *J Immunol*. 2018;201(12):3750–3758.
43. Chen X, et al. Exosomes derived from hypoxic epithelial ovarian cancer deliver microRNA-940 to induce macrophage M2 polarization. *Oncology Reports*. 2017;38(1):522–528.
44. Kolev M, et al. Inside-Out of Complement in Cancer. *Front Immunol*. 2022;13:931273.
45. Nunez-Cruz S, et al. Genetic and Pharmacologic Inhibition of Complement Impairs Endothelial Cell Function and Ablates Ovarian Cancer Neovascularization. *Neoplasia*. 2012;14(11):994-IN1.
46. Savant S, Sriramkumar S, O'Hagan H. The Role of Inflammation and Inflammatory Mediators in the Development, Progression, Metastasis, and Chemoresistance of Epithelial Ovarian Cancer. *Cancers*. 2018;10(8):251.
47. Ashton TM, et al. Oxidative Phosphorylation as an Emerging Target in Cancer Therapy. *Clinical Cancer Research*. 2018;24(11):2482–2490.

48. Anderson AS, et al. Ovarian tumor-initiating cells display a flexible metabolism. *Experimental Cell Research*. 2014;328(1):44–57.
49. Zhang W, et al. Syntaxin 6 Enhances the Progression of Epithelial Ovarian Cancer by Promoting Cancer Cell Proliferation. *Asian Pac J Cancer Prev*. 2023;24(6):2003–2010.
50. Zhang B, et al. STX5 Inhibits Hepatocellular Carcinoma Adhesion and Promotes Metastasis by Regulating the PI3K/mTOR Pathway. *J Clin Transl Hepatol*. 2023;000(000):000–000.
51. Sun H, et al. Exosomal S100A4 derived from highly metastatic hepatocellular carcinoma cells promotes metastasis by activating STAT3. *Sig Transduct Target Ther*. 2021;6(1):187.
52. Wu X, et al. Exosome-transmitted S100A4 induces immunosuppression and non-small cell lung cancer development by activating STAT3. *Clin Exp Immunol*. 2022;210(3):309–320.
53. Deo AN, et al. IGF1R- $\alpha$ 6 integrin-S100A4 network governs the organ-specific metastasis of chemoresistant epithelial ovarian cancer cells. *Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease*. 2022;1868(1):166282.
54. Link T, et al. Clinical relevance of circulating MACC 1 and S100A4 transcripts for ovarian cancer. *Molecular Oncology*. 2019;13(5):1268–1279.
55. Orsburn BC. Proteome Discoverer—A Community Enhanced Data Processing Suite for Protein Informatics. *Proteomes*. 2021;9(1):15.
56. Bonzon-Kulichenko E, et al. Revisiting Peptide Identification by High-Accuracy Mass Spectrometry: Problems Associated with the Use of Narrow Mass Precursor Windows. *J Proteome Res*. 2015;14(2):700–710.



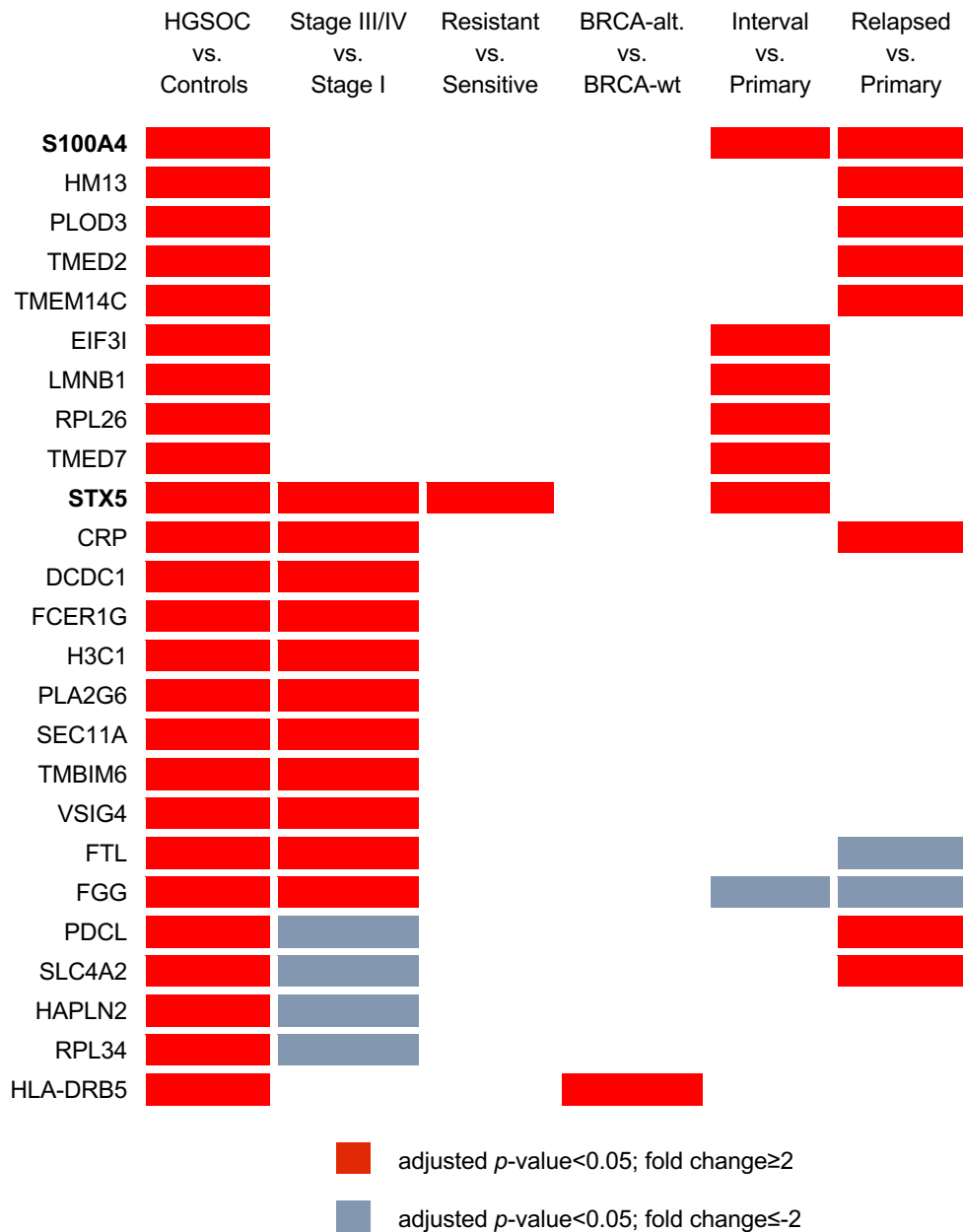
57. Navarro P, et al. General Statistical Framework for Quantitative Proteomics by Stable Isotope Labeling. *J Proteome Res.* 2014;13(3):1234–1247.
58. Wilkerson MD, Hayes DN. ConsensusClusterPlus: a class discovery tool with confidence assessments and item tracking. *Bioinformatics.* 2010;26(12):1572–1573.
59. Subramanian A, et al. Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles. *Proc Natl Acad Sci U S A.* 2005;102(43):15545–15550.
60. Benjamini Y, et al. Controlling the false discovery rate in behavior genetics research. *Behavioural Brain Research.* 2001;125(1–2):279–284.

## TABLES

**Table 1.** Demographics of the OvCa cases and controls included in the study. 'NA', Not applicable.

| Parameter               |                              | OvCa cases<br>(n=65) | Controls<br>(n=29) |
|-------------------------|------------------------------|----------------------|--------------------|
| Age                     | Mean; range                  | 61; 26-83            | 43; 25-79          |
| FIGO stage              | I-II                         | 7                    | NA                 |
|                         | III                          | 38                   |                    |
|                         | IV                           | 20                   |                    |
| Histology<br>(Cases)    | HGS                          | 54                   | NA                 |
|                         | Endometrioid                 | 6                    |                    |
|                         | LGS                          | 2                    |                    |
|                         | Clear cells                  | 2                    |                    |
|                         | Mucinous                     | 1                    |                    |
| Diagnosis<br>(Controls) | Serous ovarian cystadenoma   |                      | 7                  |
|                         | Mucinous ovarian cystadenoma |                      | 3                  |
|                         | Uterine myoma                |                      | 7                  |
|                         | Ovarian cystic teratoma      |                      | 2                  |
|                         | Endometriosis                |                      | 4                  |
|                         | Normal tissue                |                      | 2                  |
|                         | Uterine prolapse             |                      | 3                  |
| Endometrial hyperplasia |                              | 1                    |                    |
| Surgery                 | Primary                      | 35                   | NA                 |
|                         | Interval                     | 25                   |                    |
|                         | Relapse                      | 14                   |                    |
| Surgical outcome        | R0                           | 54                   | NA                 |
|                         | R1                           | 10                   |                    |
|                         | NA                           | 10                   |                    |
| BRCA status             | Altered                      | 16                   | NA                 |
|                         | Wild-type                    | 38                   |                    |
|                         | Not available                | 11                   |                    |
| BRCA 1/2 defects        | Germline                     | 7                    |                    |
|                         | Somatic                      | 6                    |                    |
|                         | Not available                | 3                    |                    |
| Platinum sensitivity    | Sensitive                    | 43                   | NA                 |
|                         | Resistant                    | 16                   |                    |
|                         | Not available                | 15                   |                    |

**Table 2.** Table depicting those 25 proteins which were differentially contained among different comparisons of interest. Red boxes represent proteins upregulated in PFD-EVs collected from experimental group (fold change>2) while blue boxes indicate proteins upregulated in the control condition.

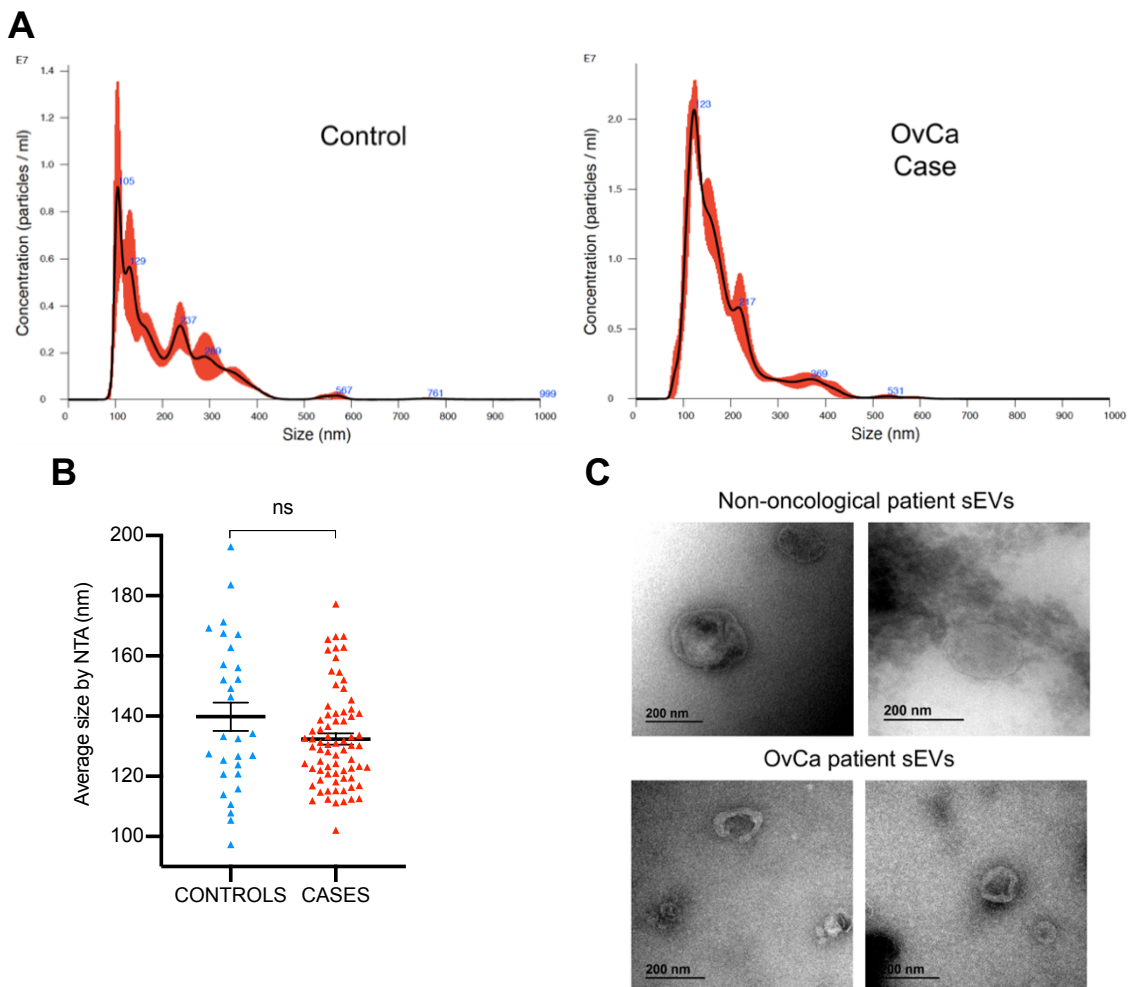


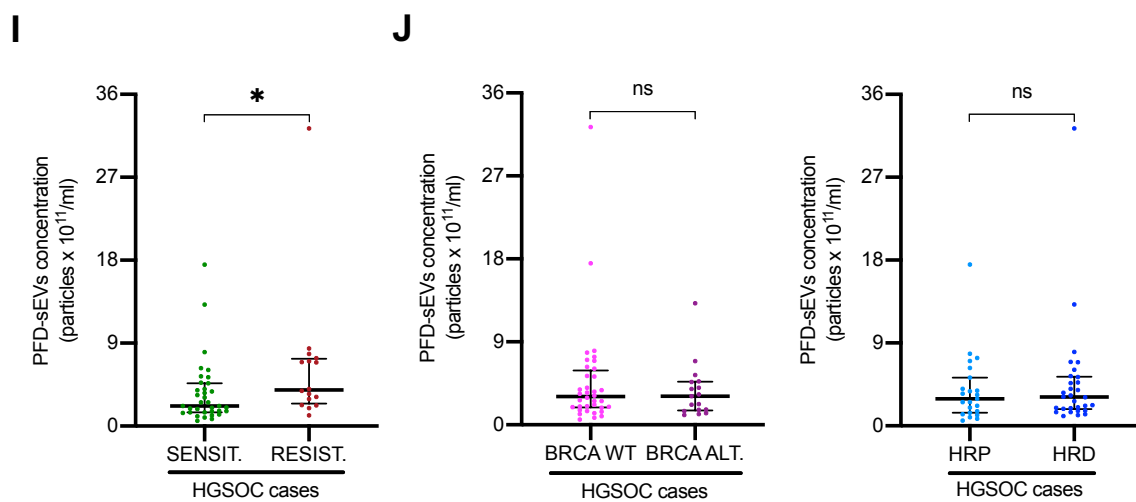
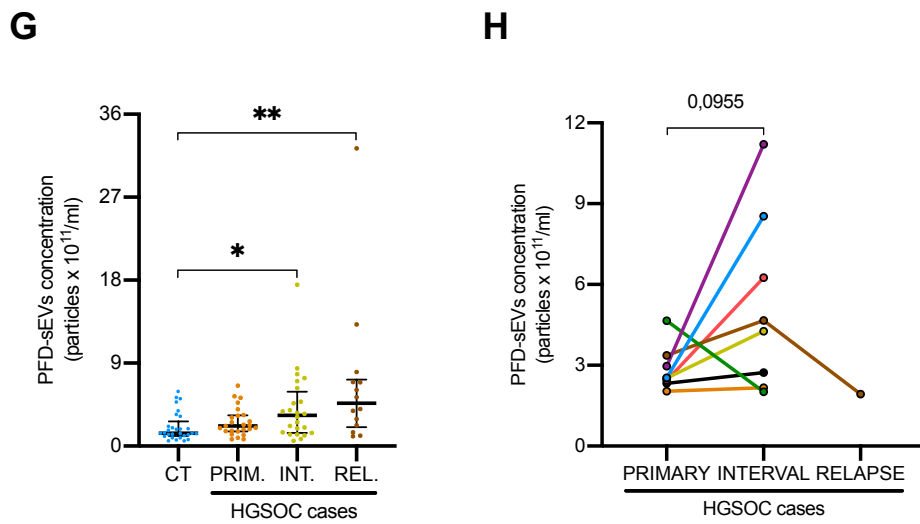
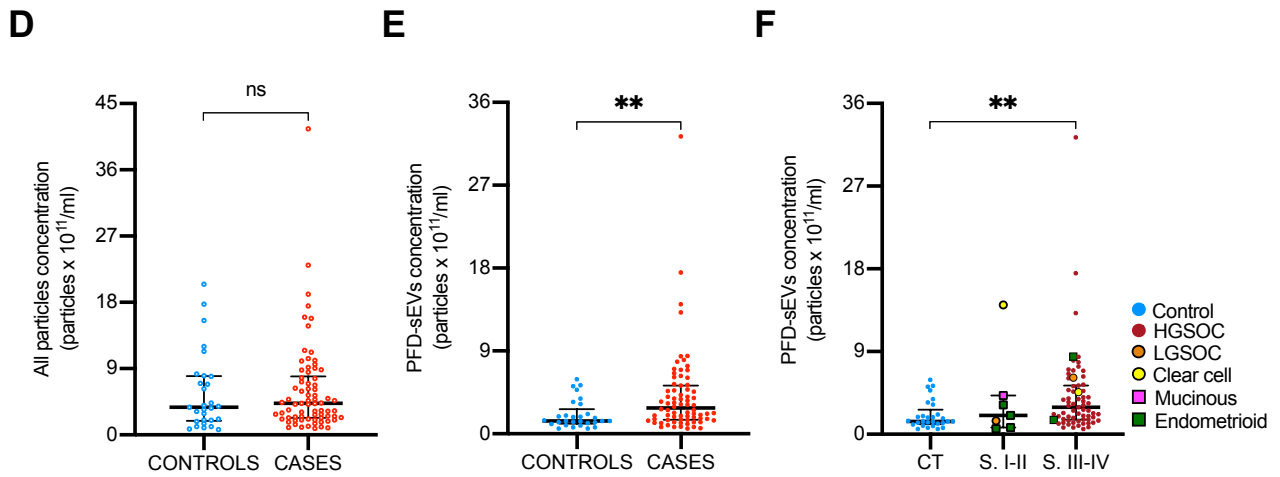
**Table 3.** 50 top differentially contained proteins in PFD-sEVs between samples belonging to cluster S-1 (n=10) vs. cluster S-2 (n=13). The left panel refers to the proteins down-contained in PFD-sEVs from the comparison of S-2 vs. S-1, where the vast majority of recurrence samples accumulated. In contrast, the right panel shows the 25 proteins over-contained in PFD-sEVs samples belonging to cluster S-2, were primary and interval surgery samples tended to cluster. logFC, fold-change logarithm; t, t statistic; adj.P.Val, adjusted *p*-value from Benjamini-Hochberg algorithm.

| Proteins over-contained in cluster S-1 |              |       |           | Proteins over-contained in cluster S-2 |             |      |           |
|--|--------------|-------|-----------|--|-------------|------|-----------|
| Protein                                | logFC        | t     | adj.P.Val | Protein                                | logFC       | t    | adj.P.Val |
| SELENBP1                               | <b>-1.76</b> | -5.66 | 4.10E-05  | LRG1                                   | <b>1.66</b> | 5.20 | 6.34E-05  |
| PRDX6                                  | <b>-1.73</b> | -5.56 | 4.10E-05  | IGKV3_20                               | <b>1.66</b> | 5.23 | 5.88E-05  |
| TUBB4B                                 | <b>-1.73</b> | -5.54 | 4.10E-05  | IGHV3_30                               | <b>1.67</b> | 5.30 | 5.53E-05  |
| UPK1B                                  | <b>-1.73</b> | -5.53 | 4.10E-05  | IGKV3D_7                               | <b>1.67</b> | 5.30 | 5.53E-05  |
| TUBA1B                                 | <b>-1.73</b> | -5.59 | 4.10E-05  | IGHG2                                  | <b>1.68</b> | 5.31 | 5.53E-05  |
| ACTB                                   | <b>-1.71</b> | -5.47 | 4.39E-05  | TECTA                                  | <b>1.68</b> | 5.19 | 6.41E-05  |
| SSR3                                   | <b>-1.68</b> | -5.27 | 5.67E-05  | IGLC2                                  | <b>1.68</b> | 5.28 | 5.67E-05  |
| ANXA1                                  | <b>-1.68</b> | -5.17 | 6.48E-05  | IGKV1D_33                              | <b>1.69</b> | 5.30 | 5.53E-05  |
| RRAS                                   | <b>-1.67</b> | -5.24 | 5.88E-05  | IGHA1                                  | <b>1.69</b> | 5.36 | 5.17E-05  |
| CLIC3                                  | <b>-1.64</b> | -5.23 | 5.88E-05  | PMEL                                   | <b>1.70</b> | 5.36 | 5.17E-05  |
| ANXA5                                  | <b>-1.64</b> | -5.12 | 7.23E-05  | CFH                                    | <b>1.70</b> | 5.39 | 4.99E-05  |
| CLIC1                                  | <b>-1.64</b> | -5.12 | 7.23E-05  | C4BPA                                  | <b>1.70</b> | 5.43 | 4.72E-05  |
| CDC42                                  | <b>-1.62</b> | -5.13 | 7.10E-05  | C4BPB                                  | <b>1.71</b> | 5.39 | 4.99E-05  |
| EIF5A                                  | <b>-1.62</b> | -4.95 | 8.74E-05  | C1S                                    | <b>1.72</b> | 5.48 | 4.39E-05  |
| ANXA7                                  | <b>-1.62</b> | -5.02 | 7.86E-05  | IGHV3_72                               | <b>1.73</b> | 5.52 | 4.10E-05  |
| DDOST                                  | <b>-1.62</b> | -5.06 | 7.65E-05  | IGLC7                                  | <b>1.73</b> | 5.55 | 4.10E-05  |
| ANXA4                                  | <b>-1.62</b> | -5.02 | 7.86E-05  | PROS1                                  | <b>1.73</b> | 5.54 | 4.10E-05  |
| GNB4                                   | <b>-1.61</b> | -5.04 | 7.86E-05  | ZNF317                                 | <b>1.74</b> | 5.58 | 4.10E-05  |
| RDX                                    | <b>-1.61</b> | -4.94 | 8.96E-05  | FCN3                                   | <b>1.76</b> | 5.65 | 4.10E-05  |
| ADH1B                                  | <b>-1.61</b> | -5.02 | 7.86E-05  | IGKC                                   | <b>1.76</b> | 5.69 | 4.10E-05  |
| IPO5                                   | <b>-1.61</b> | -4.95 | 8.74E-05  | C1R                                    | <b>1.78</b> | 5.75 | 4.10E-05  |
| PRNP                                   | <b>-1.61</b> | -5.15 | 6.81E-05  | C1QB                                   | <b>1.78</b> | 5.78 | 4.10E-05  |
| S100A6                                 | <b>-1.60</b> | -4.98 | 8.41E-05  | C1QA                                   | <b>1.78</b> | 5.83 | 4.10E-05  |
| MYL6                                   | <b>-1.60</b> | -4.94 | 8.96E-05  | IGHG1                                  | <b>1.79</b> | 5.85 | 4.10E-05  |
| MYO1D                                  | <b>-1.57</b> | -4.95 | 8.74E-05  | C1QC                                   | <b>1.79</b> | 5.85 | 4.10E-05  |

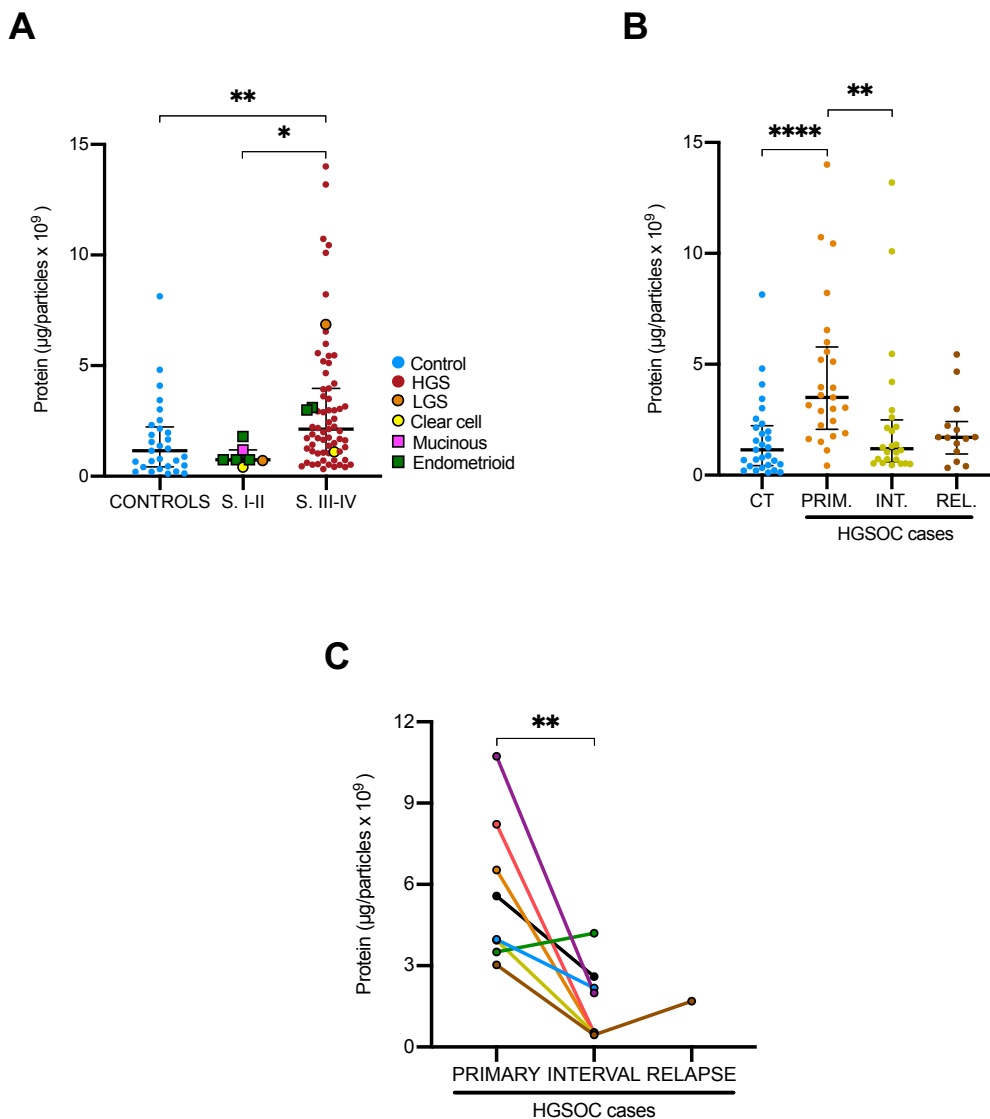
## FIGURES

**Figure 1.** Characterization of Peritoneal Fluid-Derived small Extracellular Vesicles (PFD-sEVs) in controls and OvCa cases. **(A)** Representative image of particle size distribution determined by NTA in both control and OvCa case samples. **(B)** Primary peak size of particles measured by NTA in control (n=29) and OvCa cases samples (n=74). **(C)** EV morphology observed by TEM in controls (n=2) and OvCa cases (n=2). Scale bar, 200 nm. **(D)** Concentration analysis of all particles in controls (n=29) and OvCa cases samples (n=74). **(E)** Analysis of PFD-sEVs concentration in controls (n=29) and OvCa cases samples (n=74). **(F)** Analysis of PFD-sEVs concentration in controls (CT, n=29) and OvCa cases samples (n=74) separated according to tumour stage (S., Stage I-II [n=7] or III-IV [n=67]). Additional information regarding histology was provided (HGSOC, High-Grade Serous Ovarian Carcinomas; LGSOC, Low Grade Serous Ovarian Carcinomas). **(G)** Analysis of PFD-sEVs concentration in controls (n=29) and HGSOC cases samples according to the surgical origin of the sample (PRIM., primary/diagnostic [n=25]; INT., interval [n=24]; REL., relapse [n=14]). **(H)** PFD-sEVs concentration at different time points from HGSOC cases with serial samples (n=8). **(I)** PFD-sEVs concentration according to cisplatin sensitivity (sensit., sensitive [n=35]; resist., resistant [n=16]) in HGSOC cases. **(J)** PFD-sEVs concentration in HGSOC cases according to *BRCA* status (WT, wild type [n=36] or ALT, altered [n=17]) or HRD status (HRP, homologous recombination proficient [n=23] or HRD, homologous recombination deficient [n=30]). Unless otherwise indicated, data are represented by Median and interquartile range (IQR) from each independent samples/experiments. *P*-values of significance (\*, *p*-value<0.05 and \*\*, *p*-value<0.01, Mann-Whitney test [B, D, E, I and J] or Kruskal-Wallis test with Dunn's multiple comparison test and Bonferroni adjusted *p*-values [F and G]).

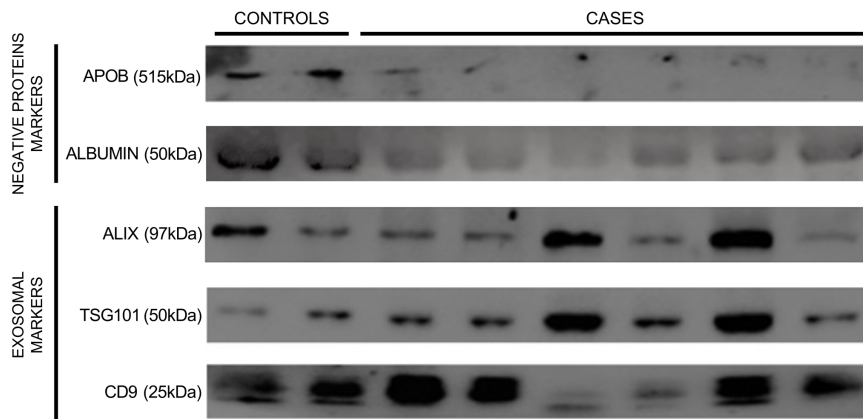




**Figure 2.** Quantitative analysis of protein content per particle per  $10^9$  in PFD-sEVs. **(A)** Ratio of protein per particle in controls (n=29) and OvCa cases samples separated according to tumour stage (Stage I-II (n=7) and III-IV (n=67)). Histological subtype information regarding each tumour is also provided. **(B)** Ratio of protein per particle in controls (CT, n=29) and HGSOc cases samples according to the surgical origin of the sample (PRIM., primary/diagnostic [n=25]; INT., interval [n=24]; REL., relapse [n=14]). **(C)** Evolution of normalized protein levels regarding the PFD-sEVs concentration in the different serial samples from HGSOc cases (n=8). **(D)** Representative Western blot of EVs markers (Alix, TSG101 and CD9) and markers of non-EVs co-isolated structures (APOB and ALB) in a selected set of controls and OvCa samples **(E)** Representative western blotting analysis of EV markers including Alix and CD9 were performed in OvCa case at different stages during the sEVs isolation process. Total PF was collected before the start of the isolation process. Supernatant sample was collected immediately after the first ultracentrifugation step and PFD-sEVs corresponds to the pellet obtained after the ultracentrifugation steps. **(F)** Quantification of normalized PAX8/CD9 ratios obtained through immunoblotting in controls (n=7) and OvCa cases samples (n=13). Unless otherwise indicated, data are represented by Median and IQR from each independent samples/experiments. *P*-values of significance (\*, *p*-value<0.05; \*\*, *p*-value<0.01 and \*\*\*\*, *p*-value<0.0001, Kruskal-Wallis test with Dunn's multiple comparison test and Bonferroni adjusted *p*-values [A and B], paired t-test [C] or Mann-Whitney test [F]).



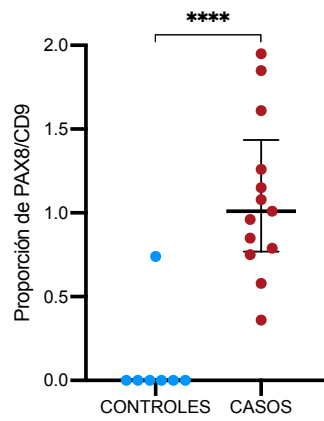
# D



# E



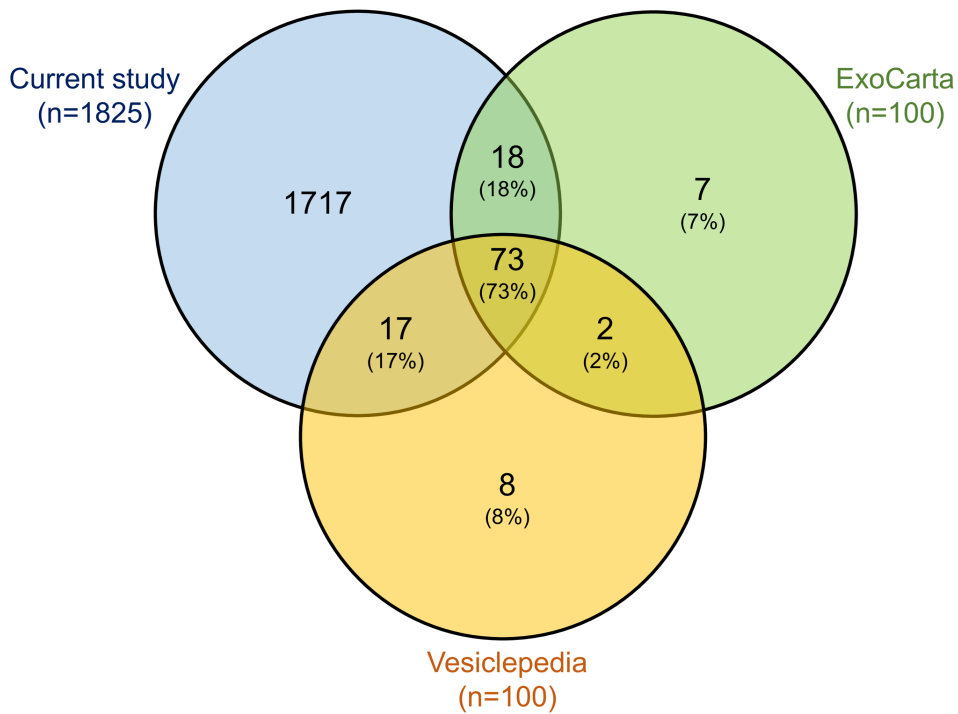
# F

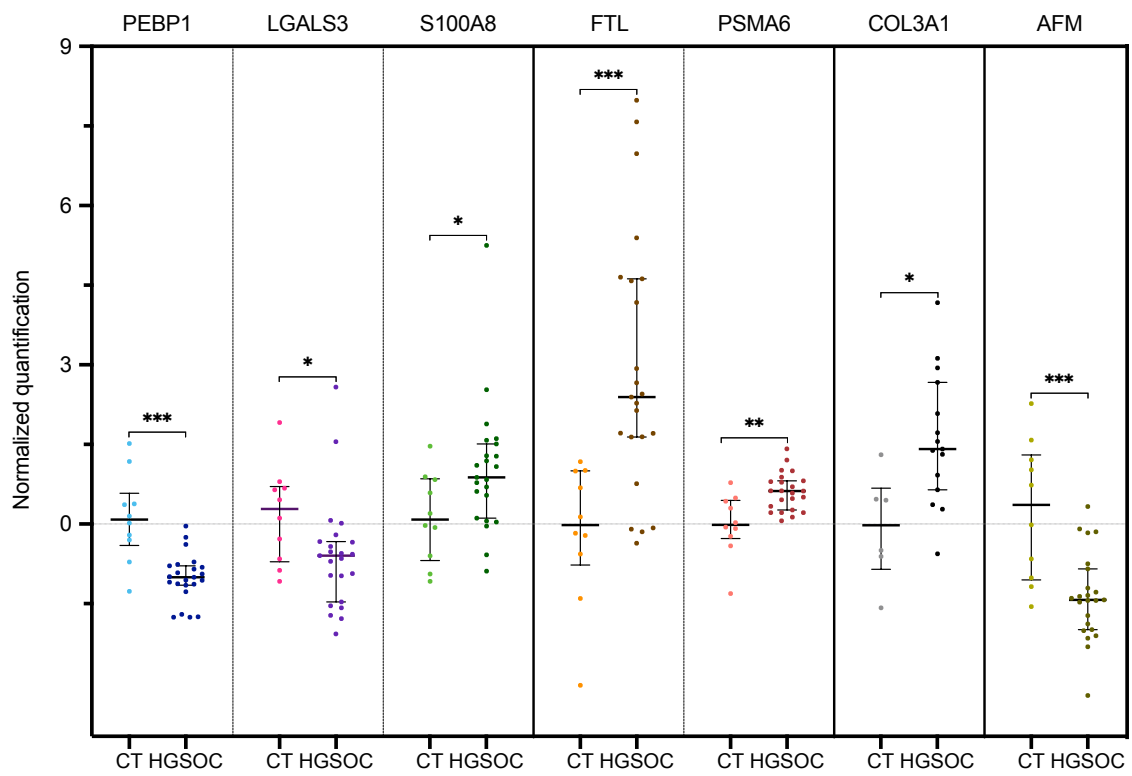
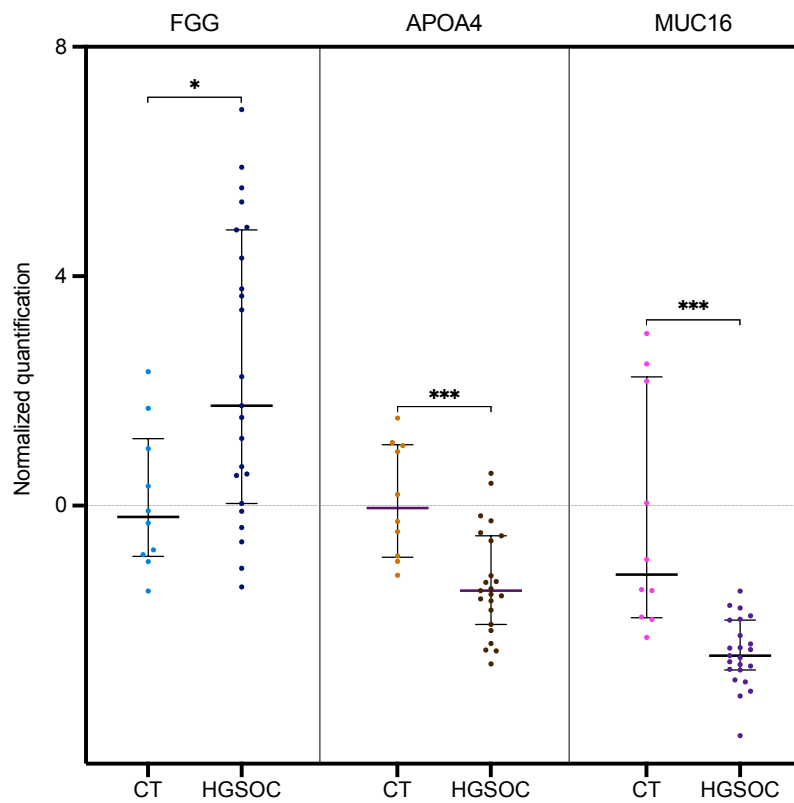




**Figure 3.** Comparison of sEVs-related protein cargo with sEVs biomarker databases and OvCa proteomic profiling from previous studies. **(A)** Venn diagrams showing the overlap between the proteins identified in PFD-sEVs in the current study and the top 100 proteins from the Vesiclepedia and ExoCarta databases. **(B)** Graph depicting the proteomic normalized quantification data for 7 proteins previously described as protein OvCa biomarkers. Comparison is established between controls (Ct, n=10) and HGSOC cases (Cases, n=23). *P*-values of significant findings (\*, *p*-value<0.05; \*\*, *p*-value<0.01 and \*\*\*, *p*-value<0.001, Mann-Whitney test) are shown in the upper part of the graph. Dark vertical lines separate markers previously described by 4 independent studies (23). **(C)** Graphs showing normalized ratios of FGG, APOA4 and MUC16 protein expression in PFD-sEVs comparing samples from our discovery cohort (HGSOC cases [n=23] vs. controls [n=10]). These factors have previously been described as diagnostic markers for OvCa in serum-derived sEVs (16). Benjamini-Hochberg adjusted *P*-values of significance (\*, *p*-value<0.05; \*\*, *p*-value<0.01 and \*\*\*, *p*-value<0.001, Mann-Whitney test).

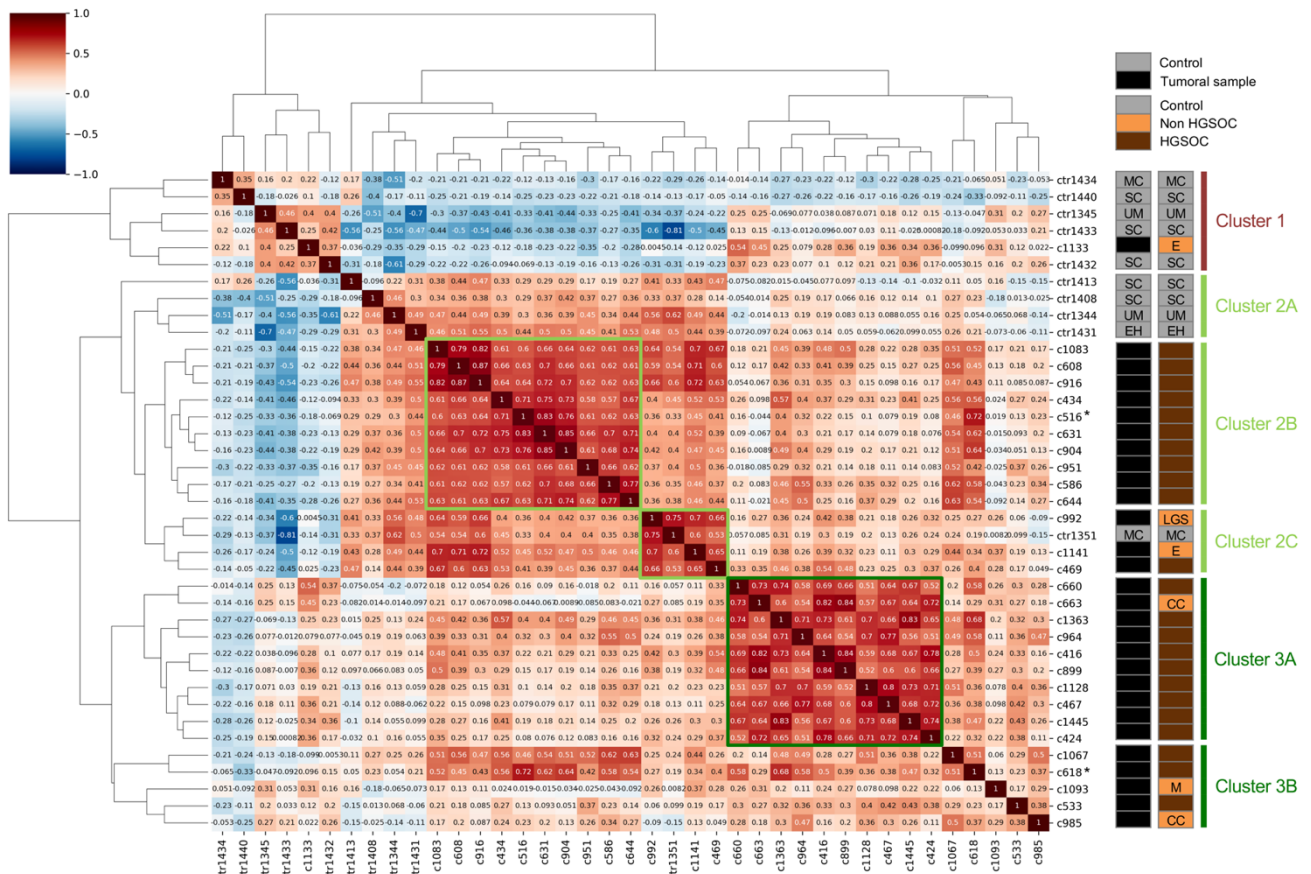
**A**



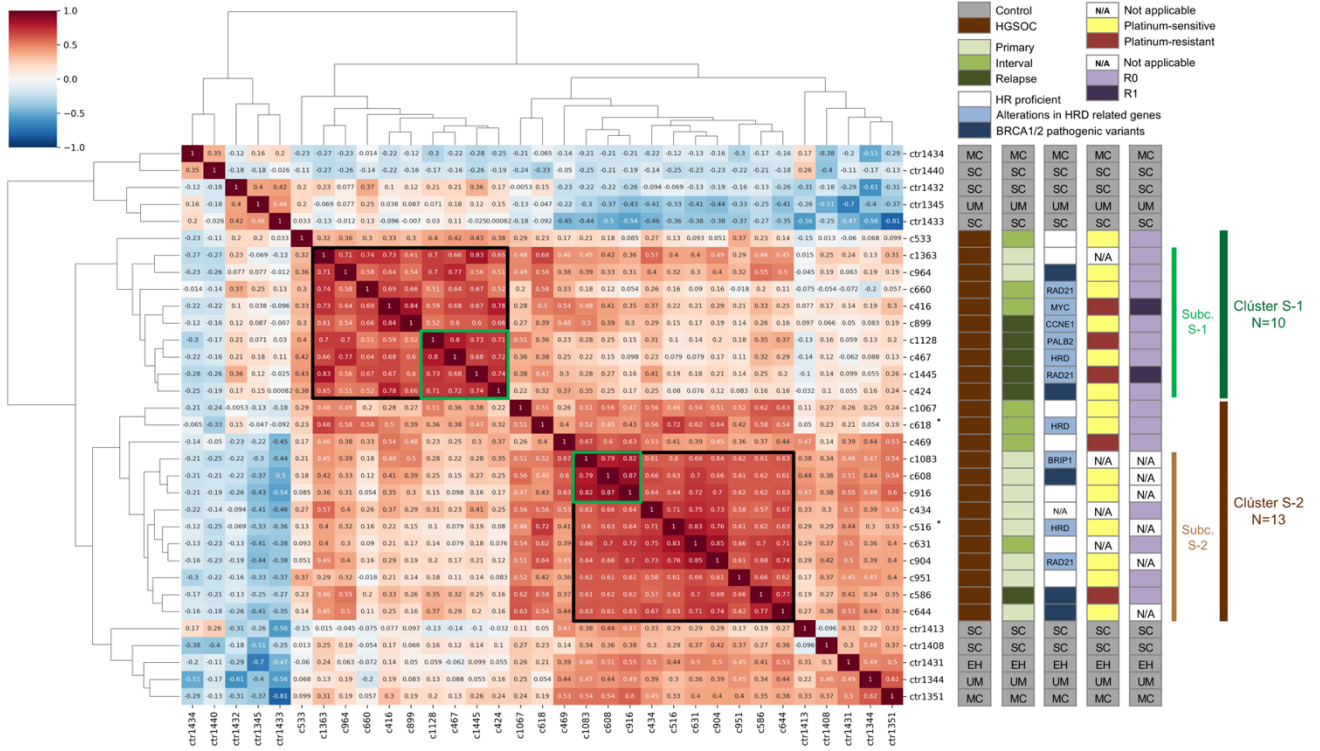
**B****C**

**Figure 4.** Correlation clustering map depicting the unsupervised analysis for the PFD-sEVs proteins profiled by mass spectrometry. **(A)** Unsupervised analysis for all cases and controls. On the right side, the main clusters (1, 2 and 3) and subclusters (2A-C and 3A-B) and the key clinical characteristics of the samples being compared (controls [SC, Serous ovarian cystadenoma; MC, Mucinous ovarian cystadenoma; UM, Uterine myoma; EH, Endometrial hyperplasia] vs. OvCa tumoral sample; HGSOC vs. Non-HGSOC [E, endometrioid; LGS, low-grade serous; M, mucinous; and CC, clear cell]) are highlighted. Correlation clusters with highest ratios are labelled in light or dark green boxes. **(B)** Unsupervised analysis for HGSOC cases and controls. On the right side, the main carcinoma clusters (S-1, n=10 and S-2, n=13) and the key clinical characteristics of the samples being compared (controls [SC, Serous ovarian cystadenoma; MC, Mucinous ovarian cystadenoma; UM, Uterine myoma; EH, Endometrial hyperplasia] vs. HGSOC; Surgical procedures: primary, interval or recurrence surgeries; HRD status: BRCA 1/2 pathogenic variants, alterations in HRD related genes, HR proficient or NA [not applicable]; Platinum sensitivity: platinum-sensitive, platinum-resistant or NA [not applicable]; Surgical outcome: R0, R1 or NA [not applicable]) are highlighted. Main correlation subclusters for both S-1 and S-2 are depicted in dark green boxes, which are mainly constituted by 55% relapses (subc.S-1, 5/9) or 80% diagnostic/primary specimens (subc.S-2, 8/10). Light green boxes include the highest correlated samples and are associated exclusively to relapses (average correlation=0.76) or diagnostic samples (average correlation, 0.82). The color bar on the left indicates the degree of correlation between two samples under study, with a value of 1 (dark red) indicating an identical sample in terms of protein cargo and -1 (dark blue) indicating potential samples with completely opposite profiles. Asterisk denotes serial samples (C516 and C618) from one HGSOC case.

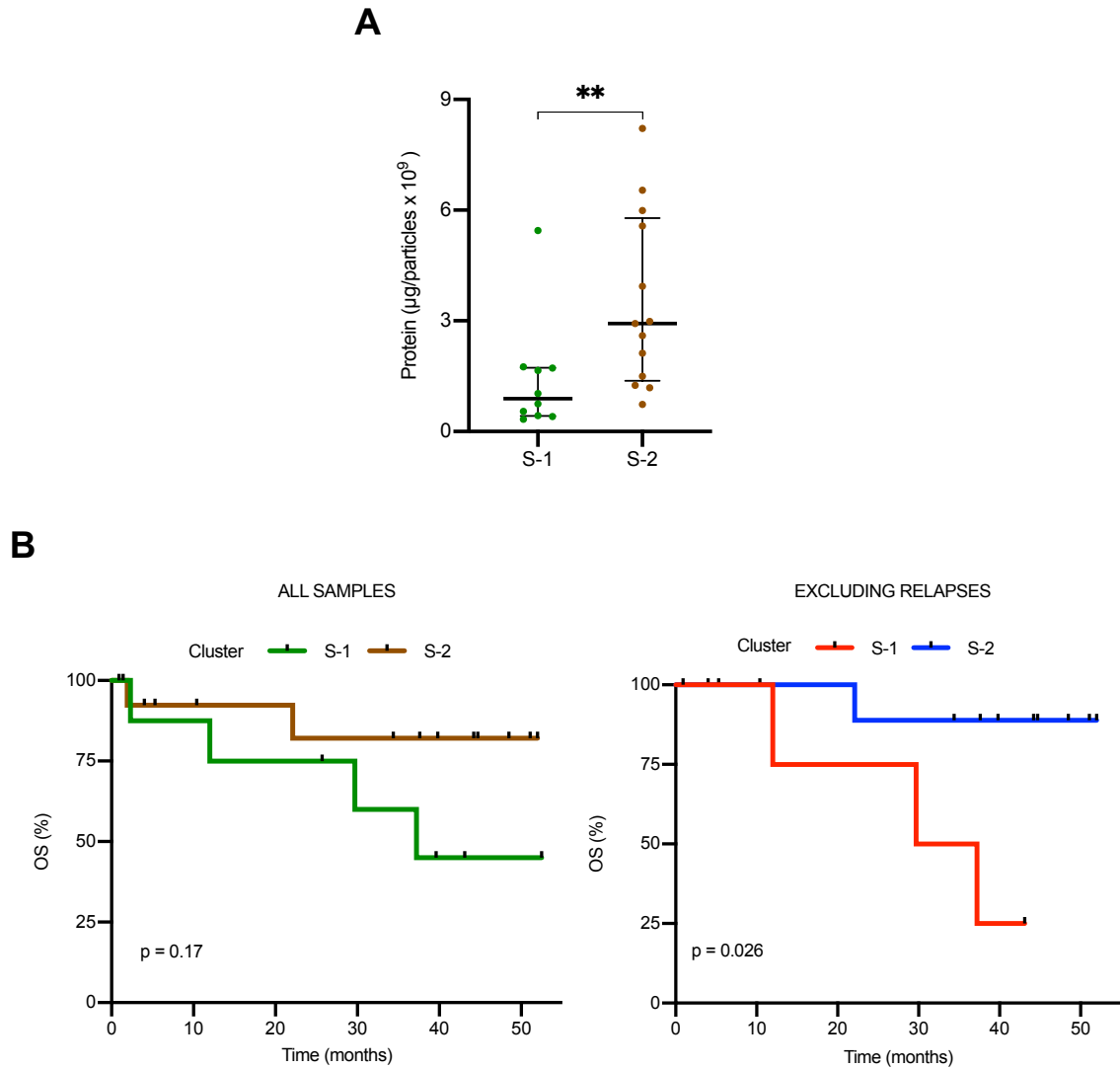
**A**



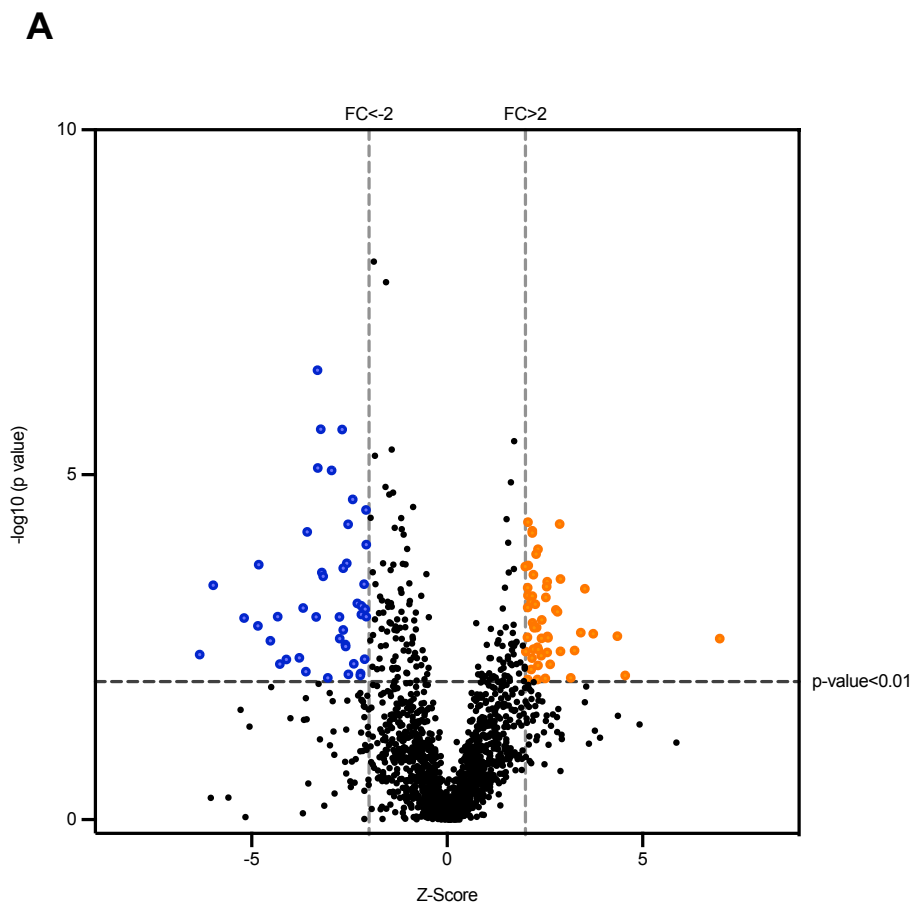
**B**

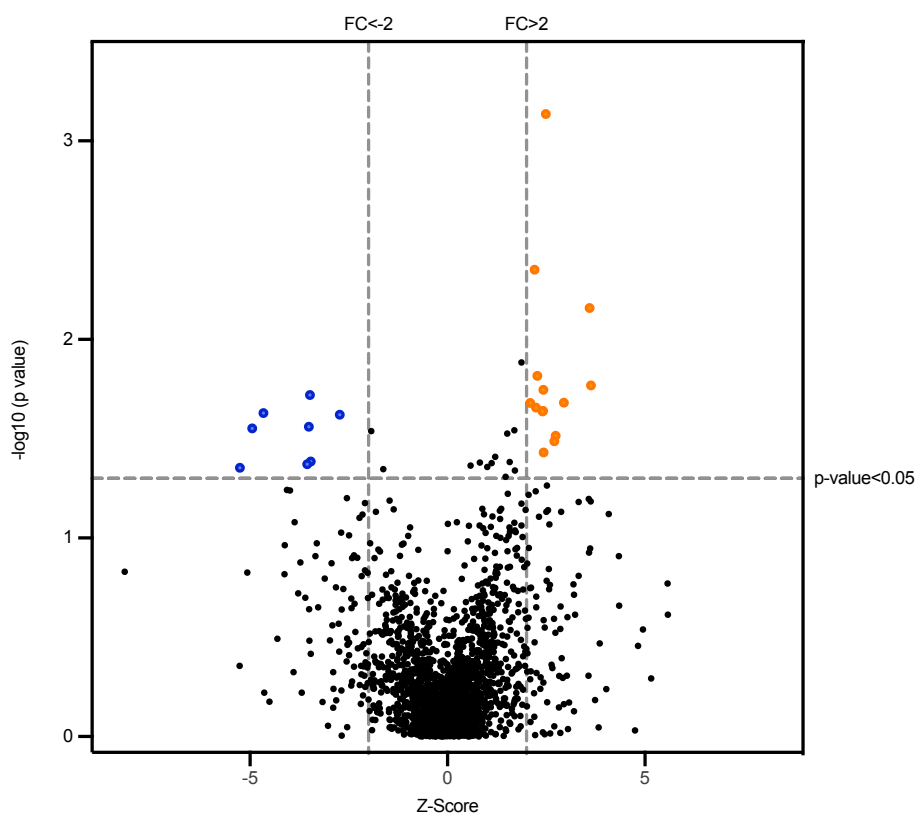
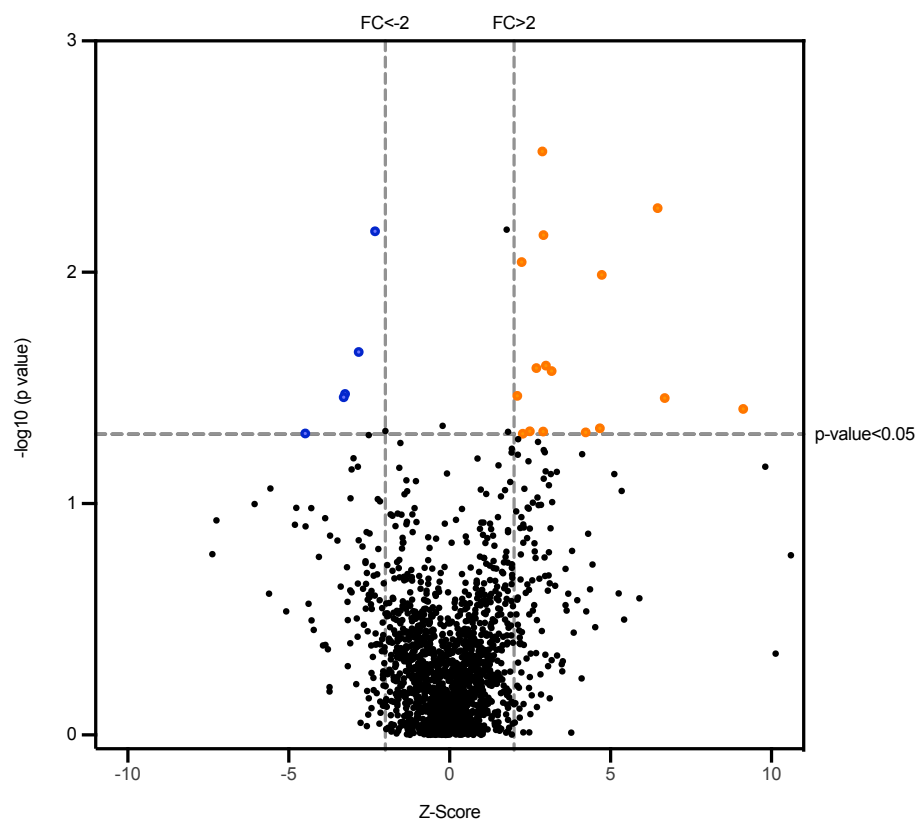


**Figure 5.** Correlation of proteomic defined HGSOC main clusters (S-1 [n=10] vs. S-2 [n=13]) with quantitative results of sEVs or with relevant clinical variables. **(A)** Association of clusters S-1 and S-2 with protein content per particle. Data are represented by Median and IQR from each independent samples/experiments. *P*-values of significant findings (\*\*, *p*-value<0.01, Mann-Whitney test). **(B)** Association of proteomic-described HGSOC clusters with overall survival (months) either including all samples (left panel; S-1 [n=10] vs. S-2 [n=13]) or excluding those patients with recurrent disease (right panel; S-1 [n=5] vs. S-2 [n=12]).

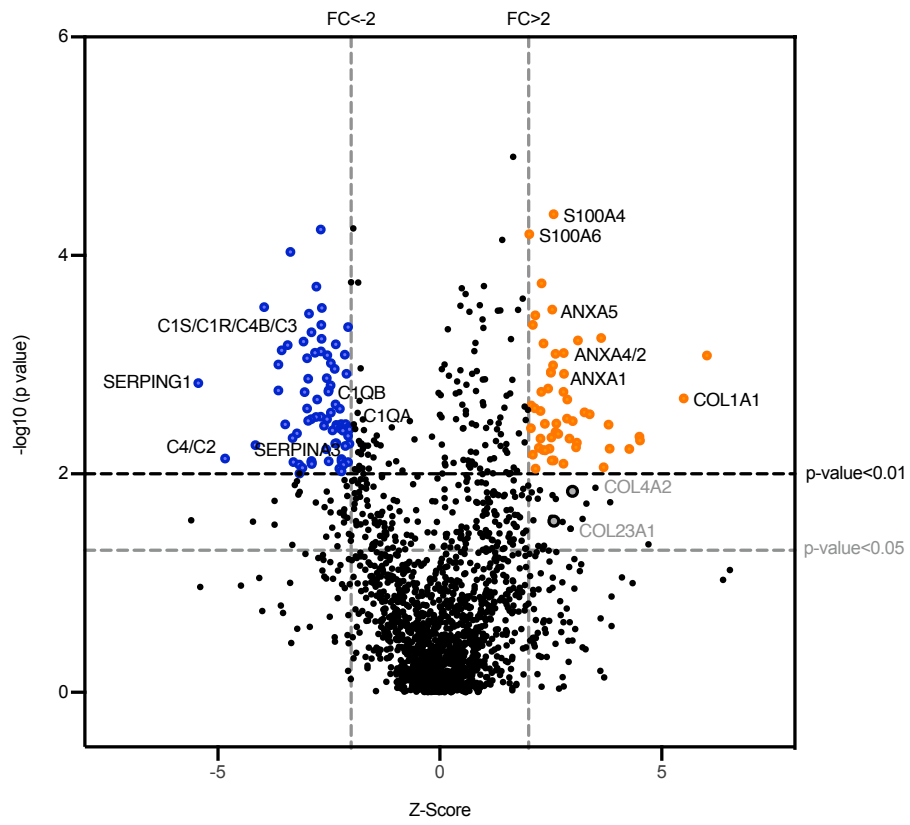


**Figure 6.** Volcano plots depicting differentially deregulated proteins (DEPs) in comparisons of interest: **(A)** HGSOC vs. controls (DEPs=96); **(B)** cisplatin resistant vs. sensitive patients (DEPs=22); **(C)** BRCA altered vs. wild type (DEPs=21) and **(D)** recurrences vs. primary (DEPs=181). Each graph depicts normalized z-scores for each of the detected master protein vs. their corresponding adjusted  $p$ -values. Dashed lines represent the adjusted  $p$ -values thresholds lower than 0.01 or 0.05 (x axis) or fold changes (FC)  $>2$  or  $<-2$  (y axis). Significantly deregulated proteins shown in the upper right quadrant (orange circles) denotes proteins overrepresented in the group under study, while proteins labelled as blue circles represents proteins overrepresented in the control group. Protein names of those functionally related to complement system or S100A/ANXA protein families are included in Figure 6D. Thresholds for significant adjusted  $p$ -values were set according to the number of samples included in each comparison.



**B****C**

**D**





**Figure 7.** Graphs depicting results related to the validation of targets of interest **(A)** STX5/CD9 or **(B)** S100A4/CD9 ratios obtained through immunoblotting in samples belonging to the validation cohort, including 5 controls and 22 HGSOC cases. Western blot bands corresponding to the above-mentioned factors were quantified using Image J software and the corresponding normalized ratio depicted in this graph as individual dots. Data are represented by median and IQR from each independent samples/experiments. *P*-values of significant findings (\*\*, *p*-value<0.01, Mann-Whitney test).

